

Basic Analytical Training

October 12, 2023

Borny Analytical Services International Consulting

Introductions

Name

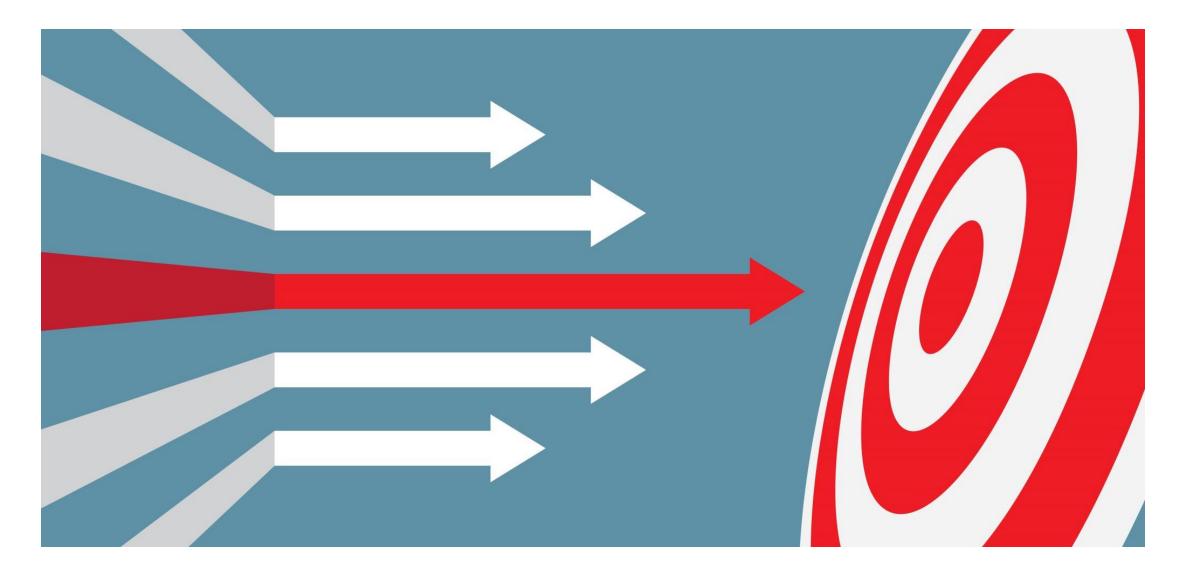
Background

Company

Type of Lab

Expectation for Workshop

Analytical Goals



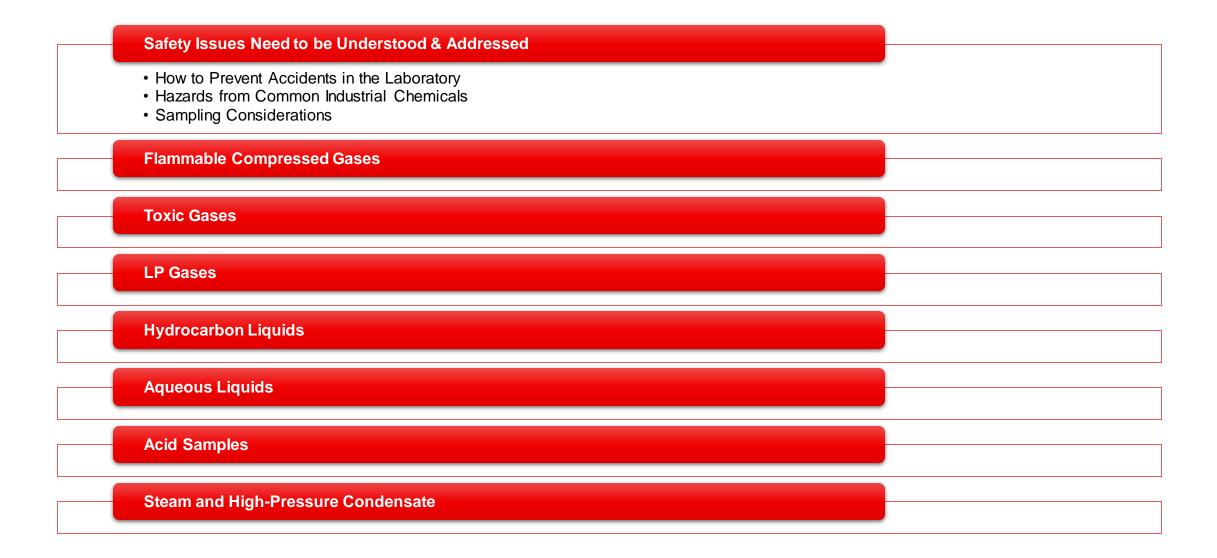
- To determine the physical, chemical, and mechanical properties of:
 - Feeds & Catalysts
 - Products & By-Products
 - Process Streams & Waste Streams
 - Other Process Materials

To Support:

- Unit Operations
- Process Control
- Troubleshooting
- Specification Requirements
- Environmental Requirements
- Determination of Contaminants

Good Analytical Measurements & Operating Data Are Essential To:

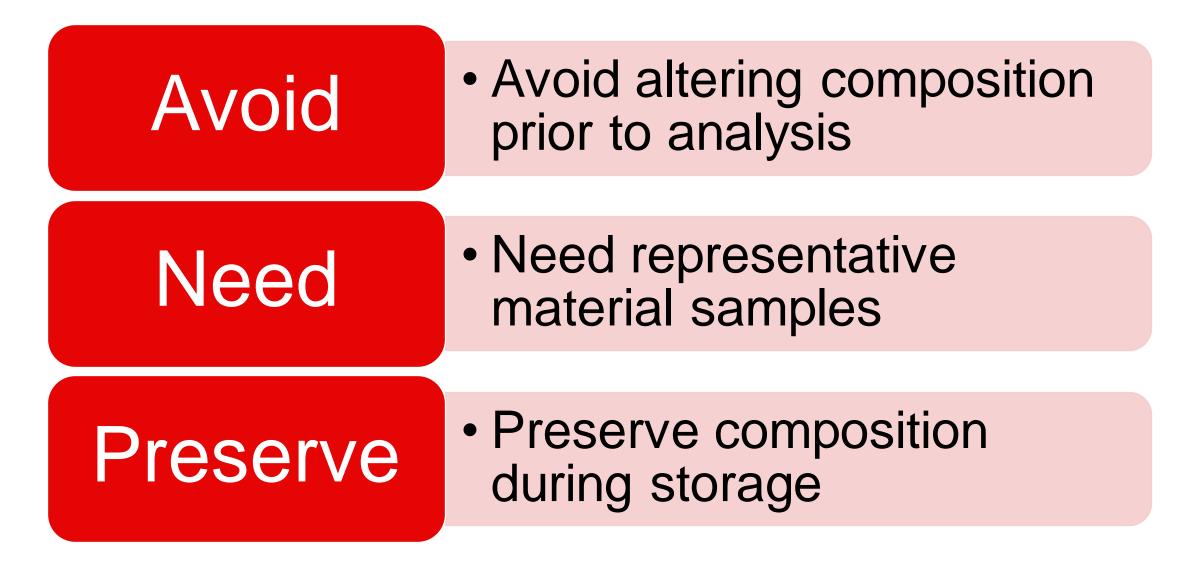
- Make critical decisions
- Achieve process goals
- Maintain product quality



Analytical General Overview



GOOD ANALYSES REQUIRE GOOD SAMPLES!

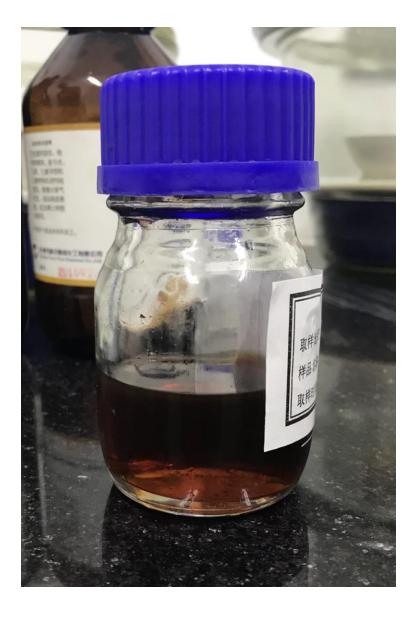


GOOD ANALYSES REQUIRE GOOD SAMPLES!

Sampling and sample handling need to be carefully considered:

- Material state (solid, gas, liquid)
- Volatile loss
- Phase separation
- Precipitation
- Temperature, pressure effects

ADDITIONAL SAMPLING REQUIREMENTS



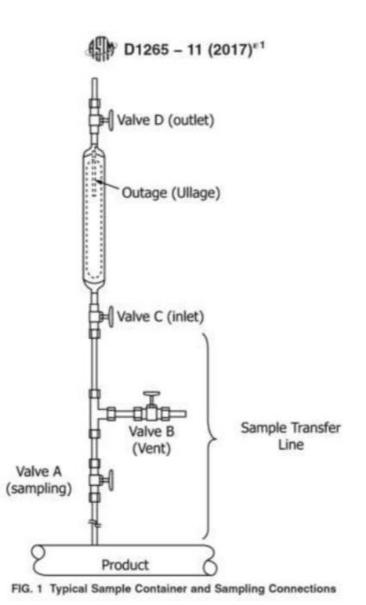
Get enough material

- Talk with the laboratory
- Present and future testing
- Future process reference

Use a proper container

- Very clean, inert, stable material
- Minimum headspace
- Dedicated

ADDITIONAL SAMPLING REQUIREMENTS



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ADDITIONAL SAMPLING REQUIREMENTS

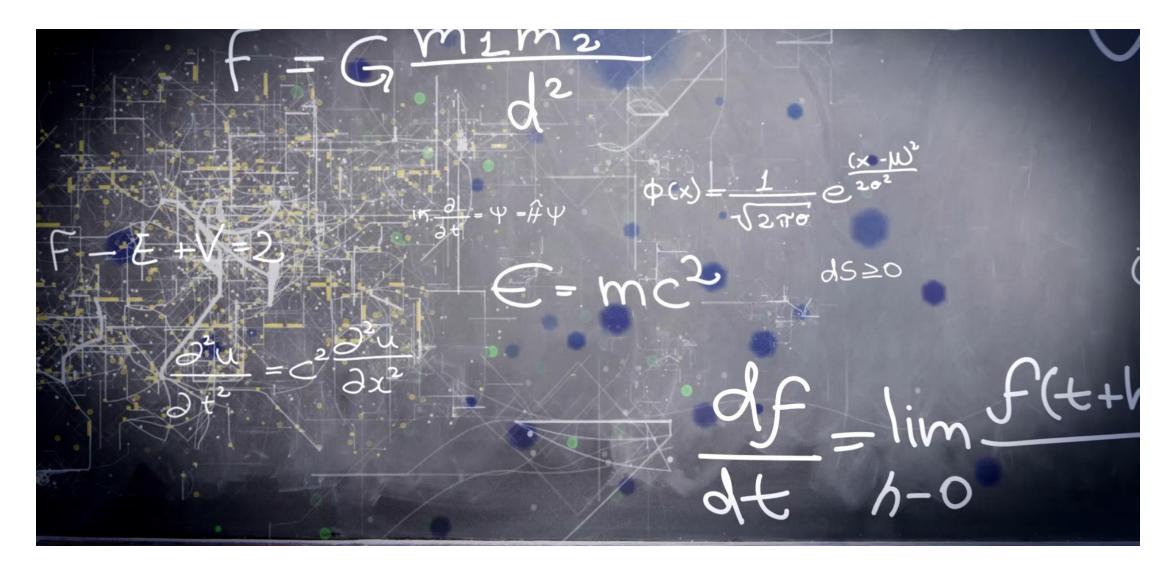


Description - Date - Time - Location

Storage

- Time Temperature Inert Atmosphere
- Save samples until results accepted or problems resolved

Analytical Methodology



GAS CHROMATOGRAPHY

Characterization of complex mixtures

Gas streams

- Petroleum liquids
- Many others

Applications cover analysis of

- Refinery gas streams
- Combustion product gases
- Inorganic gases
 Liquid streams

GAS CHROMATOGRAPHY - Broad Concentration Ranges



Trace (ppm or less)

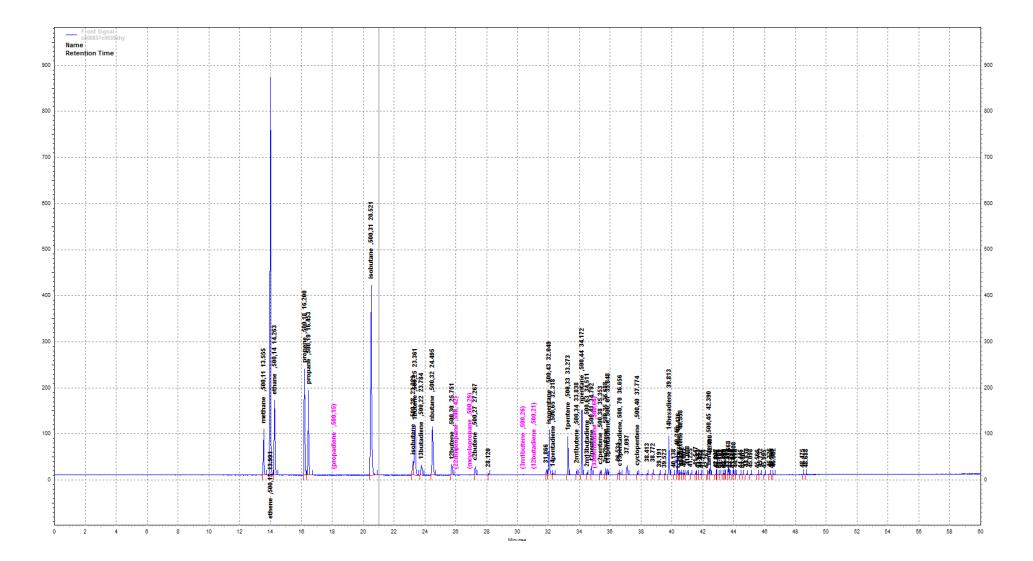
Separation Results from Vapor / Adsorbed Phase Partitioning

- First Out: Most Volatile & Least Adsorbed (lower MW, less polar)
- Last Out: Least Volatile & Most Adsorbed (higher MW, more polar)



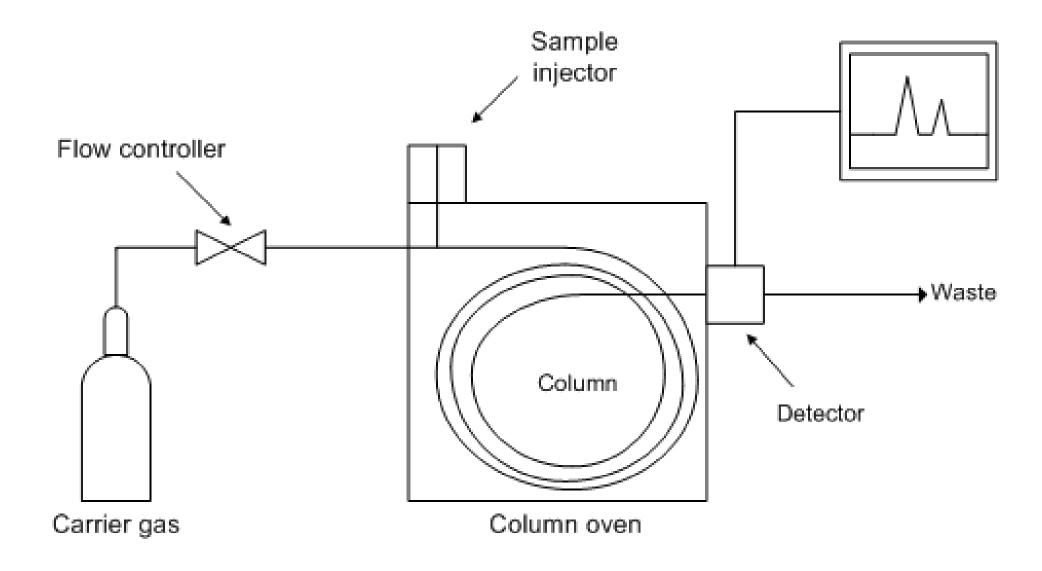
BASIC GC PRINCIPLES

Full Chromatogram

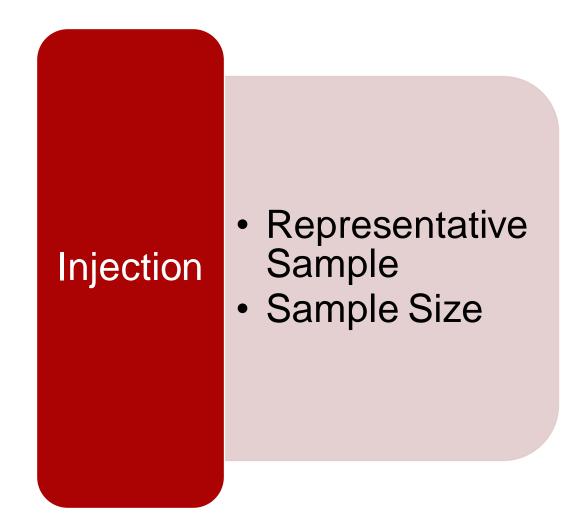


18

BASIC GC SYSTEM



GC TECHNOLOGY: THREE PRINCIPAL EVENTS





GC TECHNOLOGY: THREE PRINCIPAL EVENTS

- Resolution
- Analysis Time

Separation

Relative
 Retention
 Indices



GC TECHNOLOGY: THREE PRINCIPAL EVENTS



- Is anything eluting? What is it?
- Detection How much is
 present?
 - What is its composition?

GC TECHNOLOGY: INJECTION

TYPES of SAMPLES

- Gas sample
- Compressed Gas sample (LPG)
- Liquid Sample







GC TECHNOLOGY: INJECTION

Syringes (one shot; short hold time)

- Standard, low pressure liquids
- Special, high-pressure liquids (LPG)

Mylar Bags (many shots; intermediate hold time)

- Non-reactive, low pressure gases
- cc to Liters

Metal Cylinders (high and low pressure BUT one phase only)

- Non-condensable gases
- Liquefied gases (LPG)

Injector Valves (on- and off- line)

- Gases (cc)
- Liquids (µL)

Headspace (liquid or solid)

- Volatile components
- Purge and Trap

GC TECHNOLOGY: INJECTION

Minimum needed

- carrier gas inlet
- a septum
- septum purge
- injector insert
- heater block
- column connection

Split/Splitless

- split line or vent another set of gas lines out of the injector
- Designed so the carrier gas flow onto the column is constant
- Controls the amount of gas out the split vent to control the amount of sample to the column
- Split vent is closed Splitless injection
- Split vent is open Split injection, only a small portion of the sample to the column

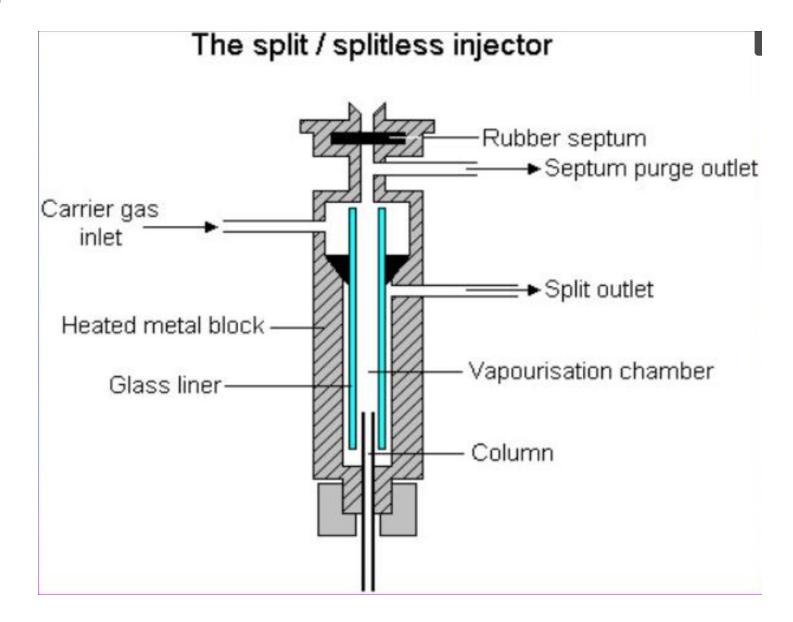
Cool on Column

- Liquid sample is drawn into the on-column syringe.
- Opening of the duck-bill septum—and stopping of carrier gas flow—is initiated by pressing down on the needle guide which splits open the duck-bill septum's "lips".
- Syringe's capillary needle is inserted down through the needle guide, through the opened duck-bill septum, completely through the cool injector into the GC column.
- Liquid sample is injected directly into a cool column ("on" the column)
- Capillary needle is withdrawn; the duck-bill septum closes when the needle guide is released; and carrier gas flow is reestablished.
- Oven's temperature program begins, and the sample vaporized to begin chromatography

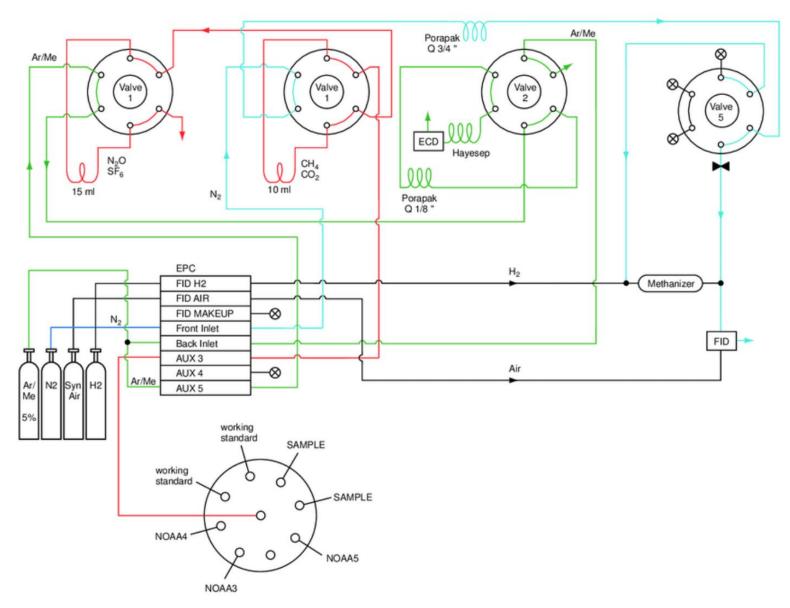
Direct on Column

- Gases (cc)
- Liquids (µL)

Split/Splitless

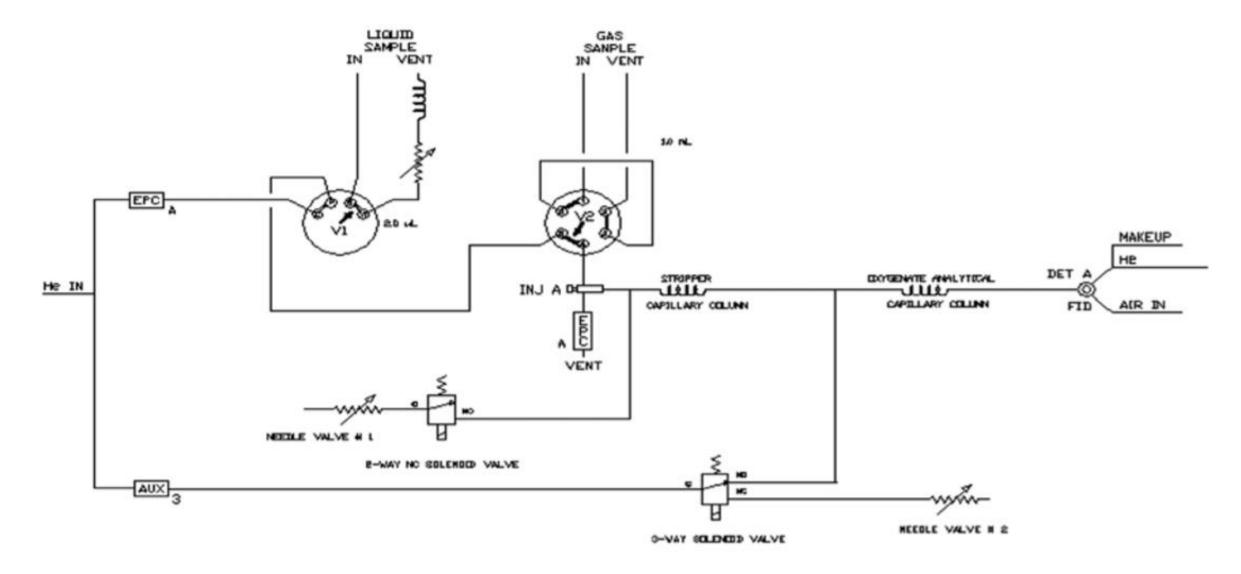


GC Valving Configuration

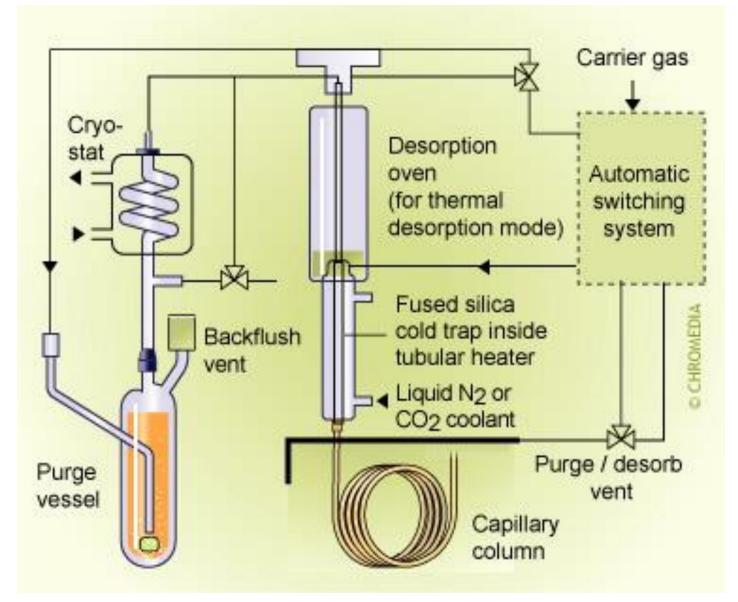


27

Schematic of a Low Level Oxygenates Multidimensional Column Configuration (Dean's Switching Method Split Inlet)



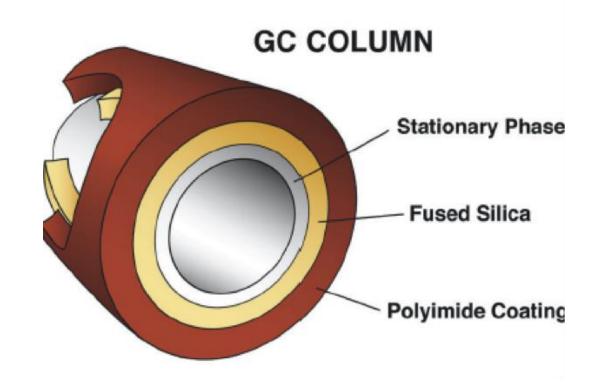
Headspace (Purge & Trap) Configuration



GC TECHNOLOGY: SEPARATION

Separation

- Boiling Point
- Carbowax
- PLOT
- Packed
- Dean Switch (multicolumn)



GC TECHNOLOGY: COLUMNS

Separation

- Kováts Retention Index
- Boiling Point
- Polar
- PLOT

GC TECHNOLOGY: COLUMNS

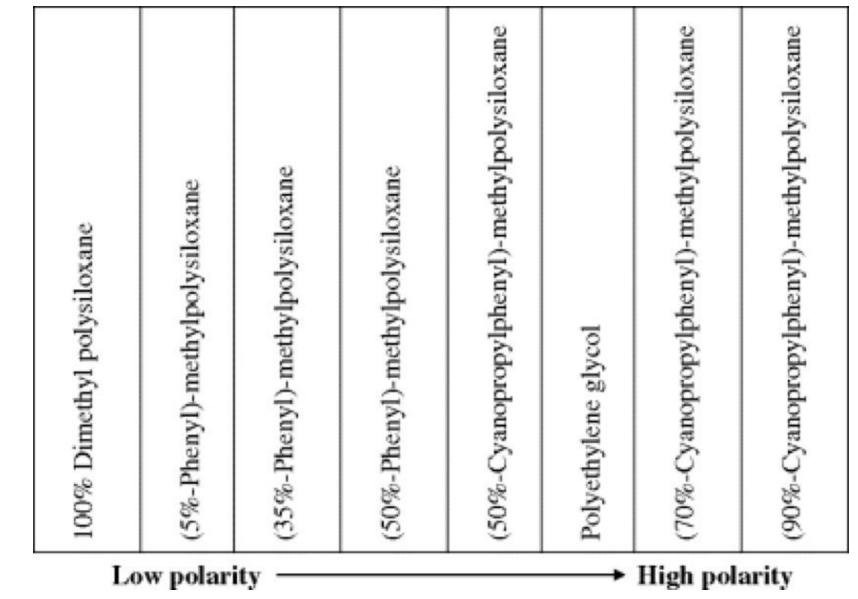
- The Kováts retention index (or Kováts index) of a compound is its retention time normalized to the retention times of adjacently eluting n-alkanes.
- It depends on the fact that log t ∝ n, where t is the retention time and n is the number of carbon atoms in the alkane.
- For an isothermal chromatogram, the following equation is used to calculate the Kováts index:

$$I = 100 \times \left[n + \frac{\log t_x - \log t_n}{\log t_{n+1} - \log t_n}\right]$$

- n is the number of carbon atoms in the n-alkane, and t is the retention time.
- Run a chromatogram of a standard alkane mixture in the range of interest
- Do a co-injection of your sample with the standard alkanes.
- Assume that the retention times were:
 - sample = 3.12 min; n-nonane = 2.71 min; n-decane = 3.89 min.
- The Kováts index for your sample is

$$I = 100 \times \left[n + \frac{\log t_x - \log t_n}{\log t_{n+1} - \log t_n} \right] = 100 \times \left[9 + \frac{\log 3.12 - \log 2.71}{\log 3.89 - \log 2.71} \right] = 939$$

Types of Columns



Types of Columns

Type of Solid Phase	Polarity	Separation Characteristics	Application	Operational Temperature Range (Approx.)
Methyl silicone DB-1, SPB-1, RTX-1	Non-polar	Boiling point order	Petroleum, solvents, high boiling point compounds	-60 to 360 °C
Phenylmethyl DB-5, RTX-5	Slightly polar Moderately polar	Phenyl groups retain aromatic compounds.	Perfumes, environmental compounds, aromatic compounds	-60 to 340 °C
Cyanopropyl phenol	Moderately polar Strongly polar	Effective at separating oxygen- containing compounds, isomers, etc.	Agricultural chemicals, PCBs, oxygen- containing compounds *Better to avoid use with FTDs (NPDs)	-20 to 280 °C
Trifluoropropyl	Moderately polar Strongly polar	Specifically retains compounds that contain halogens.	Halogen-containing compounds, polar compounds, solvents	-20 to 340 °C
Polyethylene glycol DB-Wax	Strongly polar	Strong retention of polar compounds	Polar compounds, solvents, perfumes, fatty acid methyl esters	40 to 250 °C
Porous Layer Open Tubular GS-Alumina KCI		Separation by degree of saturation	Light hydrocarbons and fixed gases	To 200 °C

GC TECHNOLOGY: DETECTION



Detection

- FID
- TCD
- SCD
- FPD/PFPD
- MS
- NCD
- TSD
- VUV
- ECD

GC TECHNOLOGY: DETECTION

Sensitivity Selectivity & Speciation

Linearity & Dynamic Range

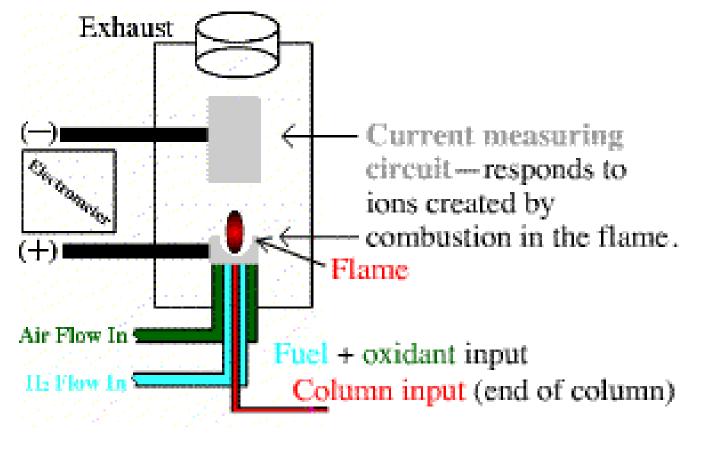
Complexity & Reliability

36

GC TECHNOLOGY: FID

- Flame Ionization
 Detector
- Carbon specific detector
- Response is $\frac{C}{MW}$
- Theoretical response
 factors possible

Flame Ionization Detector (FID)



FID Response factor calculations

 $\frac{c}{MW} = \text{calculated} = \text{normalized to C7}$ $CH4 = \frac{c}{MW} = \frac{12}{16} = 0.75 = 1.12$ $C4H8 = \frac{c}{MW} = \frac{48}{56} = 0.86 = 0.98$ $C4H6 = \frac{c}{MW} = \frac{48}{54} = 0.89 = 0.94$ $C7H16 = \frac{c}{MW} = \frac{84}{100} = 0.84 = 1.00$ $CH3OH = \frac{c}{MW} = \frac{12}{32} = 0.38 = 2.21$

Response Factors for Gas Chromatographic Analyses

by **W. A. Dietz,** Esso Research and Engineering Company Analytical Research Division, Linden, New Jersey

Abstract

While there are many types of detectors employed in gas chromatographs, most units employ either flame ionization or thermal conductivity detectors. To obtain quantitative results from the GC trace, it is necessary to use correction factors; the amount of the correction is a function of the response of a given compound to the detecting device.

Flame Ionization Detectors

Relative sensitivity values for the flame ionization detector are listed in Table I. Each area is divided by the relative sensitivity to get true area. Normalizing the results gives weight percent of each component. For hydrocarbons, with two exceptions, the values are all approximately 1.0. The two exceptions are benzene 1.12, and toluene 1.07.

For other compounds, the relative sensitivity values vary appreciably. Alcohols, for example, vary from 0.23 to 0.85; acids, from 0.01 to 0.65, etc. The use of the correct relative sensitivity is, therefore, most important when dealing with nonhydrocarbons.

Thermal Conductivity Detectors

Rosie and Grob (2) and others determined relative response values for many of the hydrocarbons and some oxygenated compounds to thermal conductivity detectors. They found the same thermal response for thermistors and hot wire filaments. Further, the response values were independent of temperature, carrier gas, flow rate, and concentration. These response values have a precision of about $\pm 3\%$. A tabulation of thermal response values are shown in Table II. These values are used as follows:

Area under the curve divided by the relative response value of that compound gives a true response value. Normalizing the true response values gives the mole percent of any component. If the sample analyzed is a gas, the normalized true response values are equal to the gas volume %.

Area under the curve multiplied by the Weight Factor gives the true weight area. When these values are normalized, the results are weight percent of each compound.

FID Response factor calculations

Table II. Effective Carbon Numbers from Published Response Data (12) Calculated Relative to Heptane								
	Factor		Factor	ECN				
Compound	(reference)	ECN	(calculated)	(theory)				
Acetylene	1.07	1.95	1.10	2				
Ethylene	1.02	2.00	1.02	2 2				
Hexene	0.99	5.82	1.02	6				
Methanol	0.23	0.52	0.222	0.5				
Ethanol	0.46	1.48	0.466	1.5				
n-Propanol	0.60	2.52	0.595	2.5				
i-Propanol	0.53	2.24	0.595(0.54)	2.5(2.25)				
n-Butanol	0.66	3.42	0.676	3.5				
Amyl alcohol	0.71	4.37	0.731	4.5				
Butanal	0.62	3.12	0.596	3				
Heptanal	0.77	6.14	0.752	6 7				
Octanal	0.78	6.99	0.782					
Capric aldehyde	0.80	8.73	0.824	9				
Acetic acid	0.24	1.01	0.238	1				
Propionic acid	0.40	2.07	0.386	2				
Butyric acid	0.48	2.95	0.487	2 3 5 6				
Hexanoic acid	0.63	5.11	0.616	5				
Heptanoic acid	0.61	5.55	0.660					
Octanoic acid	0.65	6.55	0.695	7				
Methyl acetate	0.20	1.04	0.386	2.0*				
Ethyl acetate	0.38	2.33	0.487	3.0*				
i-Propyl acetate	0.49	3.52	0.561	4.0*				
n-Butyl acetate	0.55	4.46	0.616	5.0*				
Acetone	0.49	2.00	0.49	2 3				
Methyl ethyl ketone	0.61	3.07	0.596	3				
Methyl i-butyl ketone	0.71	4.97	0.716	5				
Ethyl butyl ketone	0.71	5.66	0.752	6				
Di-i-butyl ketone	0.72	7.15	0.805	8				
Ethyl amyl ketone	0.80	7.16	0.782	7				
Cyclohexanone	0.72	4.94	0.729	5				

Journal of Chromatographic Science, Vol. 23, August, 1985

Calculation of Flame Ionization Detector Relative Response Factors Using the Effective Carbon Number Concept

James T. Scanlon and Donald E. Willis*

Corporate Research Laboratories, Monsanto Company, 800 N. Lindbergh Boulevard, St. Louis, Missouri 63167

Abstract

Equations are given for relating flame ionization detector relative response factors to the effective carbon number (ECN) of neat and derivatized components. The ECN approach can be used for calculating relative response factors in cases where pure materials are not available for detector calibration. Examples of this approach are given for the analysis of polycyclic aromatic hydrocarbons and oxygenated organics in neat form, alcohols and acids as the trimethylsilylated derivatives, and carbohydrates as the trimethylsilyl-oxime derivatives. of this paper to show how the ECN has been used for column evaluation, as a check on experimentally determined response factors for neat and derivatized compounds, and for the calculation of response factors for compounds which cannot be obtained in pure form.

Experimental

All of the data were obtained using Model 3700 gas chromatographs (Varian) and a Model 3353-E lab automation system (Hewlett Bookord). Two columns were used. The first was a

This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.



Designation: D5441 – 98 (Reapproved 2017)

Standard Test Method for Analysis of Methyl Tert-Butyl Ether (MTBE) by Gas Chromatography¹

This standard is issued under the fixed designation D5441; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers the determination of the purity of methyl tert-butyl ether (MTBE) by gas chromatography. It also provides a procedure to measure impurities in MTBE such as C_4 to C_{12} olefins, methyl, isopropyl and tert-butyl alcohols, methyl sec-butyl and methyl tert-amyl ethers, acetone, and methyl ethyl ketone. Impurities are determined to a minimum concentration of 0.02 mass %.

1.2 This test method is not applicable to the determination of MTBE in gasoline.

1.3 Water cannot be determined by this test method and must be measured by a procedure such as Test Method D1364 and the result used to normalize the chromatographic values.

1.4 A majority of the impurities in MTBE is resolved by the test method, however, some co-elution is encountered.

1.5 This test method is inappropriate for impurities that boil

mendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.

2. Referenced Documents

- 2.1 ASTM Standards:²
- D1364 Test Method for Water in Volatile Solvents (Karl Fischer Reagent Titration Method)
- D3700 Practice for Obtaining LPG Samples Using a Floating Piston Cylinder
- D4057 Practice for Manual Sampling of Petroleum and Petroleum Products
- D4307 Practice for Preparation of Liquid Blends for Use as Analytical Standards
- D4626 Practice for Calculation of Gas Chromatographic Response Factors
- E355 Practice for Gas Chromatography Terms and Relationships
- E594 Practice for Testing Flame Ionization Detectors Used

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		Components			
		Retention Time m, min		Typical Response	Density at
No. Component	50	100	150	Factor	approximately 20 °C g /mL
1 Methanol ^D	3.72	7.84	12.89	3.20	0.7914
2 Isobutylene ^E	3.85	8.00	13.39	1.18	0.5942
3 Butane	3.92	8.08	13.59	1.17	0.5788
4 Trans-2-butene	3.99	8.16	13.77	1.13	0.6042
5 Cis-2-butene	4.10	8.29	14.11	1.10	0.6213
6 3-methyl-1-butene	4.41	8.67	14.95	1.05	0.6272
7 Acetone	4.61	8.91	15.29	1.85	0.7899
8 Isopentane	4.66	8.93	15.51	1.04	0.6201
9 2-propanol	4.77	9.06	15.69	1.88	0.7855
10 1-pentene	4.82	9.15	15.95	1.05	0.6405
11 2-methyl-1-butene	4.95	9.31	16.15	1.00	0.6504
12 Pentane	5.00	9.37	16.37	1.05	0.6262
13 Trans-2-pentene	5.12	9.49	16.61	1.05	0.6482
14 Tert-butanol	5.20	9.57	16.70	1.30	0.7887
15 Cis-2-pentene	5.26	9.67	16.94	1.05	0.6556
16 2-methyl-2-butene	5.37	9.78	17.13	1.00	0.6623
17 Cyclopentene	6.17	10.72	18.84	1.00	0.7457
18 Methyl tert-butyl ether	6.51	11.11	19.15	1.53	0.7405
19 2,3-dimethyl-1-butene	6.55	11.17	19.25	1.00	0.6803
20 4-methyl-cis-2-pentene	6.57	11.21	19.36	1.00	0.669
21 2-methylpentane	6.63	11.28	19.39	1.00	0.6532
22 Methyl ethyl ketone	6.86	11.48	19.65	1.51	0.8054
23 3-methylpentane	7.09	11.80	20.17	1.00	0.6645
24 Sec-butyl methyl ether	7.22	11.93	20.23	1.53	0.7415
25 Ethyl tert-butyl ether	8.54	13.36	21.85	1.50	0.7519
26 Tert-amyl methyl ether	11.93	16.27	25.19	1.41	0.7703
27 3,5-dimethyl-1-hexene	14.85	18.23	27.39	0.90	0.708
28 2,4,4-trimethyl-1-pentene	15.03	18.40	27.65	0.90	0.715
29 2,4,4-trimethyl-2-pentene	16.17	19.27	28.47	0.90	0.7218
30 3,4,4-trimethyl-trans-2-pentene	17.86	20.86	30.19	0.90	0.739
31 2,3,4-trimethyl-2-pentene	19.02	22.00	31.28	0.90	0.7434
32 4,4-dimethyl-2-neopentyl-1-pentene	26.26	30.67	41.33	0.90	0.759
33 2,2,4,6,6-pentamethyl-3-heptene	26.46	30.92	41.64	0.90	0.759

TABLE 2 Typical Retention Times on Three Columns, Relative Mass Response Factors^A and Densities^{B,C} for Common MTBE Product

^A Response factors are relative to heptane = 1.00. ^B See Driesbach.¹²

^C See Weast.¹¹

^D Methanol coelutes with isobutane on the 50 m and 100 m columns but is separated on the 150 m column. Subambient temperature conditions will separate these compounds.

^E Isobutylene and 1-butene co-elute on all three columns at the typical temperature conditions. These components are known to separate using subambient temperature.

Carbon No.	Saturated Paraffins	Unsaturated Paraffins	Saturated Naphthenes	Unsaturated Naphthenes	Aromatics
1	1.1207	-	-	-	-
2	1.0503	-	-	-	-
3	1.0268	0.9799	-	-	-
4	1.0151	0.9799	-	-	-
5	1.0080	0.9799	0.9799	0.9517	-
6	1.0034	0.9799	0.9799	0.9564	0.9095
7	1.0000	0.9799	0.9799	0.9598	0.9195
8	0.9975	0.9799	0.9799	0.9623	0.9271
9	0.9955	0.9799	0.9799	0.9642	0.9329
10	0.9940	0.9799	0.9799	0.9658	0.9376
11	0.9927	0.9799	0.9799	0.9671	0.9415
12	0.9916	0.9799	0.9799	0.9681	0.9447
13	0.9907	0.9799	0.9799	0.9690	0.9474
14	0.9899	0.9799	0.9799	0.9698	0.9497
15	0.9893	0.9799	0.9799	0.9705	0.9517

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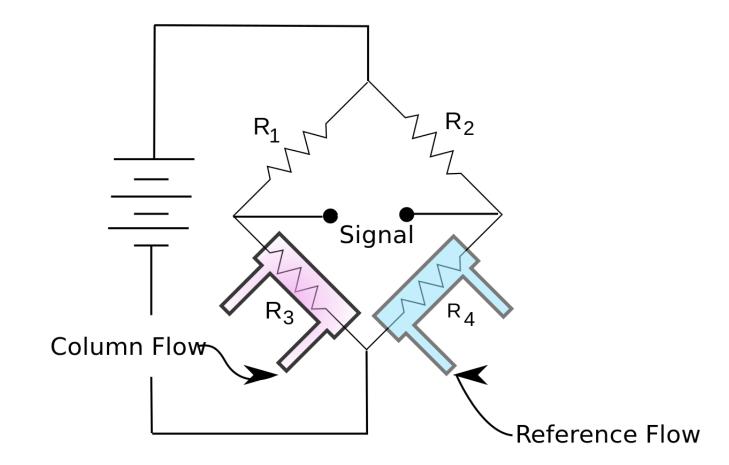
D6730 – 01 (2016)

	Oxygenates Relative Response Factors								
								Auto/Oil	
	Lab 1	Lab 2	Lab 3	Lab 4	Ave.	Std. Dev.	%SD	RRF	
Methanol	3.0760	3.0477	2.9779	2.9230	3.0062	0.0691	2.30	3.0965	
Ethanol	2.1888	2.0797	2.1755	2.0640	2.1270	0.0642	3.02	2.0953	
t-Butanol	1.2975	1.3189	1.3312	1.2989	1.3116	0.0163	1.24	1.3368	
MTBE	1.5279	1.5590	1.4860	1.5024	1.5188	0.0318	2.09	1.5016	
ETBE	1.3848	1.3720	1.3804	1.3720	1.3773	0.0064	0.46	1.4032	
TAME	1.3383	1.2993	1.3598	1.3340	1.3329	0.0250	1.88	1.3775	

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GC TECHNOLOGY: TCD

- Thermal Conductivity Detector
- Universal Detector
- Response is based on reference filament
- External standard needed



TCD Response factor

Table II. Response Factors for Thermal Conductivity Detectors.

B.P. °C	Compound	MW	Thermal Response	Weight Factor
			•	
Normal	Paraffins (2,3)			
- 161	Methane	16	35.7	0.45
- 89	Ethane	30	51.2	0.59
- 42	Propane	44	64.5	0.68
- 0.5	Butane	58	85	0.68
+ 36	Pentane	72	105	0.69
68	Hexane	86	123	0.70
98	Heptane	100	143	0.70
126	Octane	114	160	0.71
151	Nonane	128	177	0.72
174	Decane	142	199	0.71
196	Undecane	156	198	0.79
254	Tetradecane	198	234	0.85
	C_{20} - C_{36}	—		0.72
Branche	d Paraffins (2,3)			
-12	Isobutane	58	82	0.710
+ 28	Isopentane	72	102	0.707
10	Neopentane	72	99	0.727
50	2,2-Dimethylbutane	86	116	0.741
58	2,3-Dimethylbutane	86	116	0.741
60	2-Methylpentane	86	120	0.714
63	3-Methylpentane	86	119	0.725
79	2,2-Dimethylketone	100	133	0.752
81	2,4-Dimethylpentane	100	129	0.775
90	2,3-Dimethylpentane	100	135	0.741

Response Factors for Gas Chromatographic Analyses

by **W. A. Dietz,** Esso Research and Engineering Company Analytical Research Division, Linden, New Jersey

Abstract

While there are many types of detectors employed in gas chromatographs, most units employ either flame ionization or thermal conductivity detectors. To obtain quantitative results from the GC trace, it is necessary to use correction factors; the amount of the correction is a function of the response of a given compound to the detecting device.

Flame Ionization Detectors

Relative sensitivity values for the flame ionization detector are listed in Table I. Each area is divided by the relative sensitivity to get true area. Normalizing the results gives weight percent of each component. For hydrocarbons, with two exceptions, the values are all approximately 1.0. The two exceptions are benzene 1.12, and toluene 1.07.

For other compounds, the relative sensitivity values vary appreciably. Alcohols, for example, vary from 0.23 to 0.85; acids, from 0.01 to 0.65, etc. The use of the correct relative sensitivity is, therefore, most important when dealing with nonhydrocarbons.

Thermal Conductivity Detectors

Rosie and Grob (2) and others determined relative response values for many of the hydrocarbons and some oxygenated compounds to thermal conductivity detectors. They found the same thermal response for thermistors and hot wire filaments. Further, the response values were independent of temperature, carrier gas, flow rate, and concentration. These response values have a precision of about $\pm 3\%$. A tabulation of thermal response values are shown in Table II. These values are used as follows:

Area under the curve divided by the relative response value of that compound gives a true response value. Normalizing the true response values gives the mole percent of any component. If the sample analyzed is a gas, the normalized true response values are equal to the gas volume %.

Area under the curve multiplied by the Weight Factor gives the true weight area. When these values are normalized, the results are weight percent of each compound.

TCD Response factor

Table II. (continued)

B.P. °C	Compound	MW	Thermal Response	Weight Factor
Cyclopa	raffins			
49	Cyclopentane	70	97	0,720
72	Methylcyclopentane	84	115	0.730
88	1,1-Dimethylcyclopentane	98	124	0.787
103	Ethylcyclopentane	98	126	0.775
100	cis-1,2-Dimethylcyclopentane	98	125	0.780
91	cis + trans-1,3-Dimethylcyclopentane	98	125	0.780
116	1.2.4-Trimethylcyclopentane (CTC)	112	136	0.825
109	1,2,4-Trimethylcyclopentane (CCT)	112	143	0.783
81	Cyclohexane	84	114	0.735
101	Methylcyclohexane	98	120	0.820
120	1,1-Dimethylcyclohexane	112	141	0.794
119-12	41,4-Dimethylcyclohexane	112	146	0.769
132	Ethylcyclohexane	112	145	0.775
155	n-Propylcyclohexane	126	158	0.800
139	1,1,3-Trimethylcyclohexane	126	139	0.907
Inorgan	ic Compounds			
	Argon	40	42	0.95
	Nitrogen	28	42	0.67
	Oxygen	32	40	0.80
	Carbon dioxide	44	48	0.915
	Carbon monoxide	28	42	0.67
	Carbon tetrachloride	154	108	1.43
	Iron carbonyl ($Fe(CO_s)$)	195	150	1.30
	Hydrogen sulfide	34	38	0.89
	Water	18	33	0.55
Hetero	Compounds			
131	Pyrrole	67	86	0.780
132	Hexylamine	101	104	0.970
11	Ethyleneoxide	44	58	0.758
35	Propyleneoxide	58	80	0.730
- 62	Hydrogen sulfide	34	38	0.890
7	Methyl mercaptan	48	59	0.810
35	Ethyl mercaptan	62	87	0.720
68	1-Propanethiol	76	101	0.750
66	Tetrahydrofuran	72	83	0.870
119	Thiophane (cyclic sulfide)	88	103	0.855
165	Ethyl silicate	208	208	0.995
21	Acetaldehyde	44	65	0.680

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Abstract

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Flame Ionization Detectors

Relative sensitivity values for the flame ionization detector are listed in Table I. Each area is divided by the relative sensitivity to get true area. Normalizing the results gives weight percent of each component. For hydrocarbons, with two exceptions, the values are all approximately 1.0. The two exceptions are benzene 1.12, and toluene 1.07. For other compounds, the relative sensitivity values vary appreciably. Alcohols, for example, vary from 0.23 to 0.85; acids, from 0.01 to 0.65, etc. The use of the correct relative sensitivity is, therefore, most important when dealing with nonhydrocarbons.

Thermal Conductivity Detectors

Rosie and Grob (2) and others determined relative response values for many of the hydrocarbons and some oxygenated compounds to thermal conductivity detectors. They found the same thermal response for thermistors and hot wire filaments. Further, the response values were independent of temperature, carrier gas, flow rate, and concentration. These response values have a precision of about $\pm 3\%$. A tabulation of thermal response values are shown in Table II. These values are used as follows:

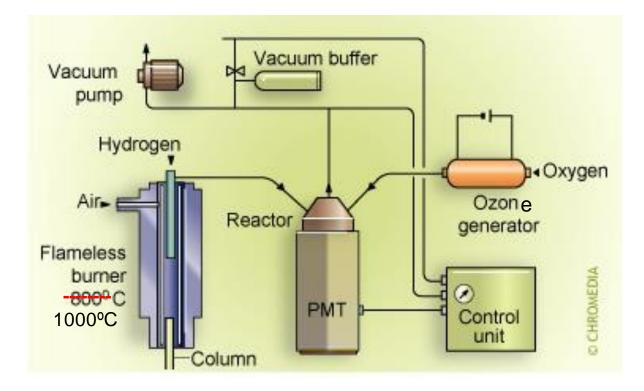
Area under the curve divided by the relative response value of that compound gives a true response value. Normalizing the true response values gives the mole percent of any component. If the sample analyzed is a gas, the normalized true response values are equal to the gas volume %.

Area under the curve multiplied by the Weight Factor gives the true weight area. When these values are normalized, the results are weight percent of each compound.

46

GC TECHNOLOGY: SCD

- Sulfur Chemiluminescence Detector
- Sulfur specific detector
- Response is equimolar for all sulfur
- External Standard calibration needed, but only one sulfur-containing compound necessary
- $R S + O_2 \rightarrow SO_2 + CO_2 + H_2O$
- $SO_2 + H_2 \rightarrow H_2S$
- $H_2S + O_3 \rightarrow SO_2 *$
- $SO_2 * \rightarrow SO_2 + h\gamma$

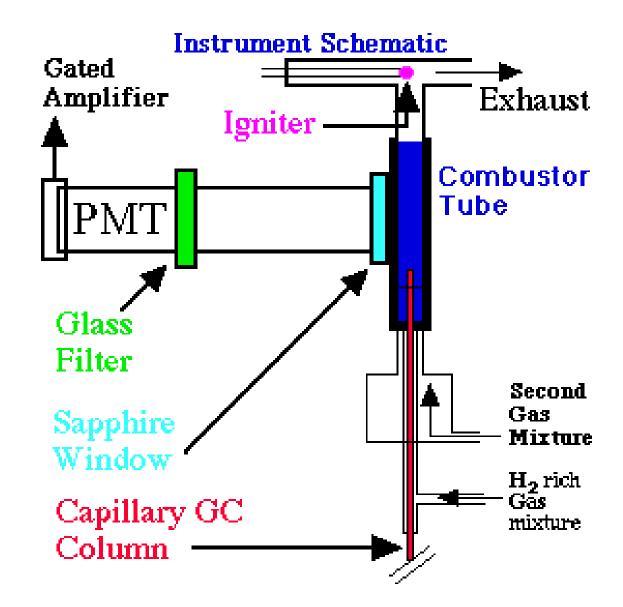


GC TECHNOLOGY: FPD/PFPD

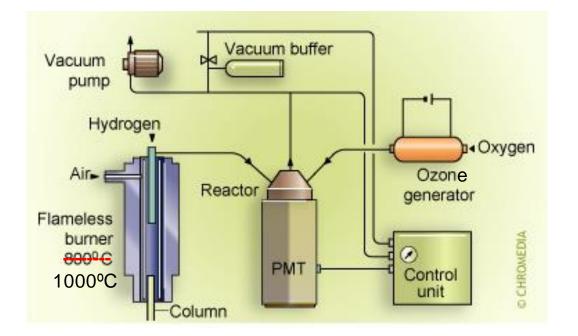
- Pulsed Flame Photometric
 Detector
 - Multielement specific detector, up to 28 S, P, N, As, Se, Sn, Ge, Ga, Sb, Te, Br, Cu, In, Mn, Fe, Ru, Rh, Cr, Ni, Eu, V, W, B, Si, Al, Pb, Bi and C

٠

- Response is proportionally related to the square root of the mass
- External Standard Needed



GC TECHNOLOGY: NCD



- Nitrogen Chemiluminescence Detector
- Nitrogen specific detector
- Response is equimolar for all nitrogen
- External standard needed but only one nitrogen-containing compound necessary

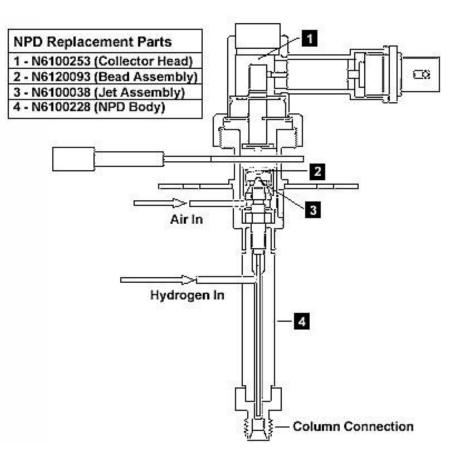
•
$$R - N + O_2 + H_2^{Opt} \rightarrow NO + CO_2 + H_2O$$

•
$$NO + O_3 \rightarrow NO_2 *$$

• $NO_2 * \rightarrow NO_2 + h\gamma$

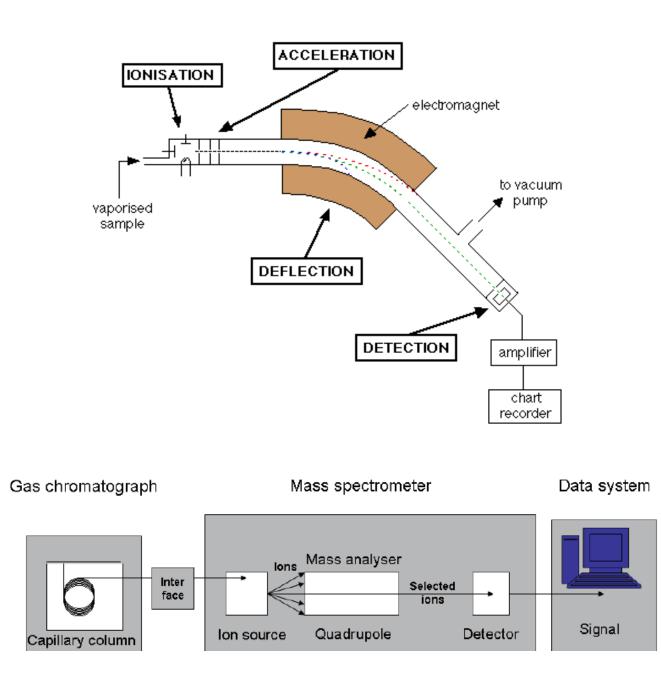
GC TECHNOLOGY: TSD/NPD

- Thermionic Specific Detector
- Nitrogen and
 Phosphorous specific detector
- Response is based on ionization



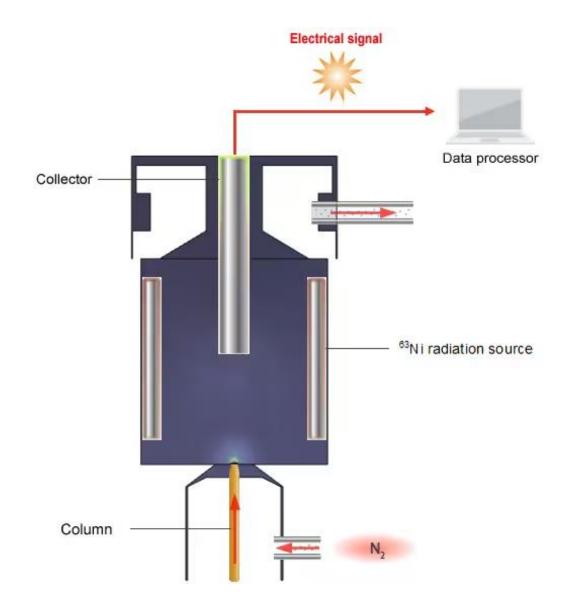
GC TECHNOLOGY: MS

- Mass Spectrometer Detector
- Mass selective detector
- Response is based on ionization
- Stage 1: Ionization
- Stage 2: Acceleration:
- Stage 3: Deflection:
- Stage 4: Detection



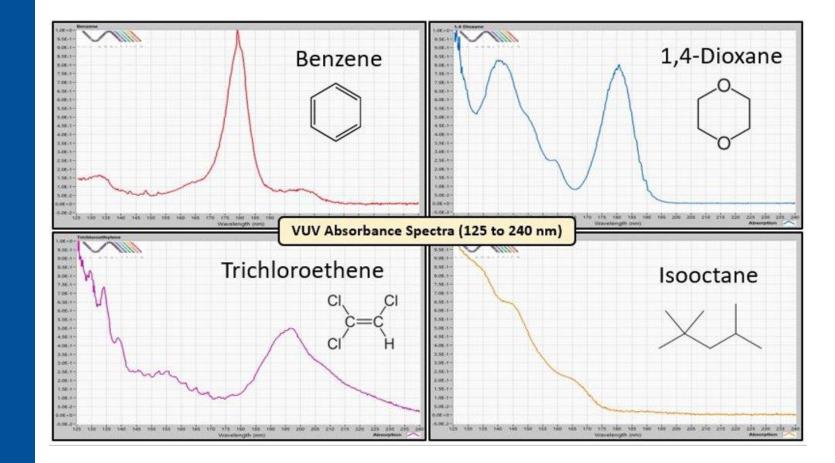
GC TECHNOLOGY: ECD

- Electron Capture Detector
- Detect organic halogen compounds
- Has a radioactive source
- Compound of interest absorbs electron and reduce the number of electrons the collectors sees



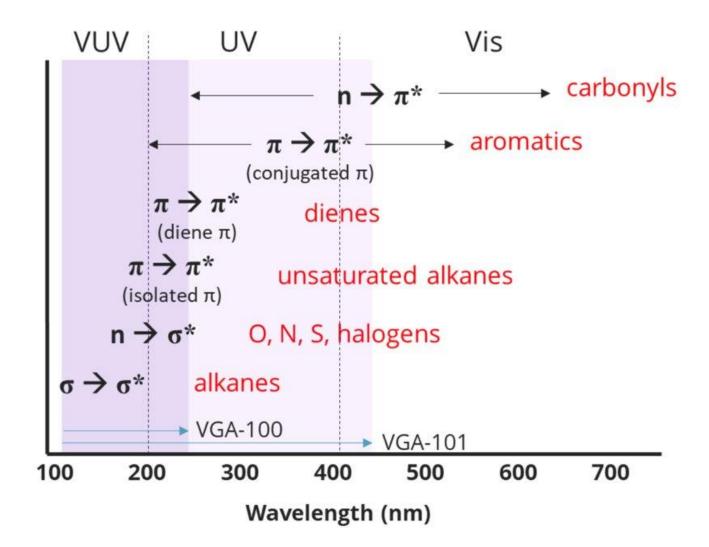
GC TECHNOLOGY: VUV

- Vacuum Ultraviolet
 Detector
- Nearly Universal Detector



GC TECHNOLOGY: VUV

- Vacuum Ultraviolet
 Detector
- Nearly Universal Detector



GC TECHNOLOGY: OTHER DETECTORS

BID Barrier Discharge Ionization Detector

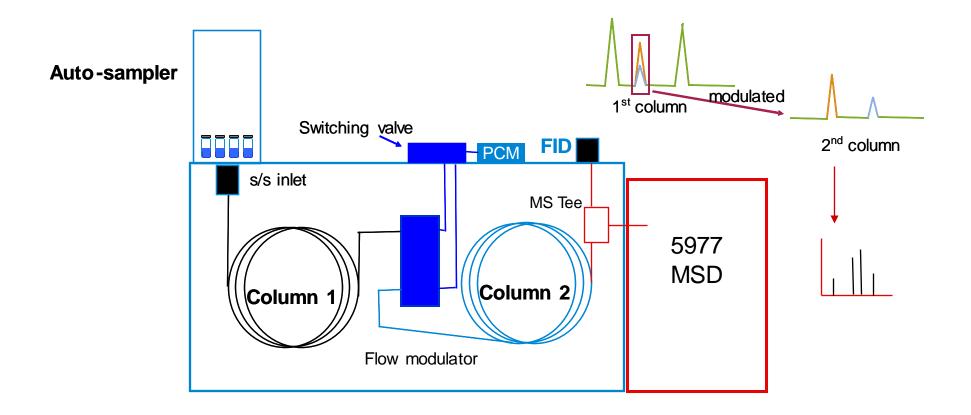
HID Helium Discharge Ionization Detector

PDD Pulsed Discharge Detector

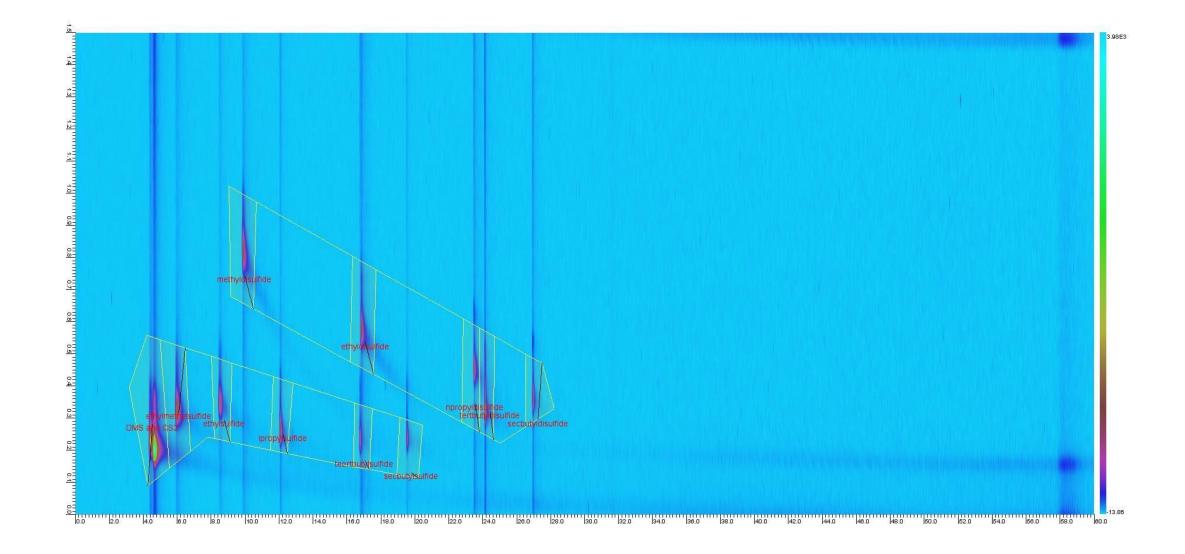
GC TECHNOLOGY

GC x GC

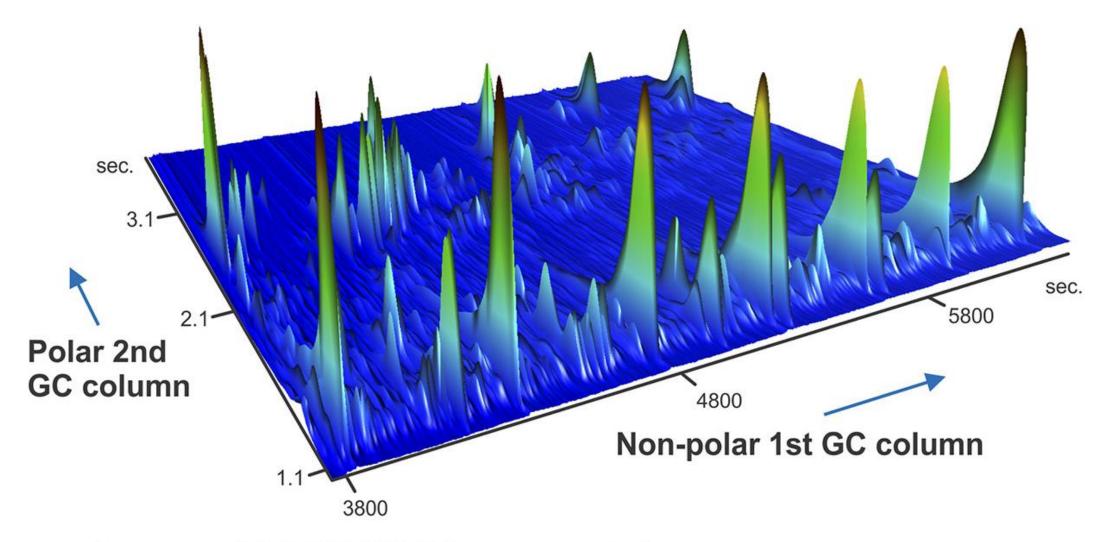
Basic system layout for GCxGC FID/MSD



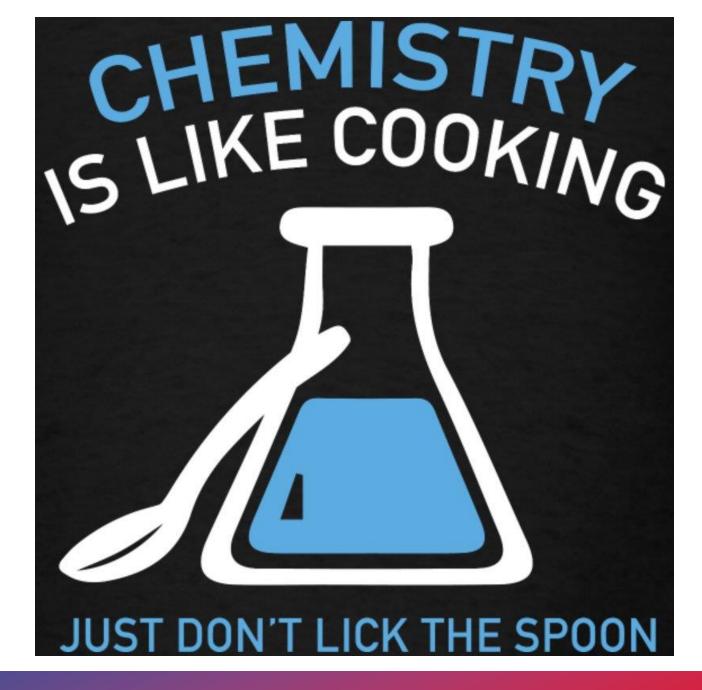
GC x GC - SCD



58



The power of GC×GC-TOFMS for your samples!



GC TECHNOLOGY: CALCULATIONS

Normalization

 Using theoretical response factor or calculated response factors, the total is equal to 100%

External Standard

 Peak area is compared to a known standard and used to calculate a response factor

Internal Standard

 A know amount of non-interfering compound is added to the sample and is used to calculate the amount of the components

GC TECHNOLOGY: INTERNAL STANDARD (ISTD)

Since the ratio of concentration between an ISTD and an analyte remains constant, even when instrument drift or sample is lost during treatment, the ratio of concentrations (analyte concentration / ISTD concentration) remain the constant.



To use the ISTD method:

Solutions of known concentration of the analyte are prepared

To each of those standards a known and similar amount of the internal standard solution

Similar amount of the internal standard is added to the samples too

Instrumental signal of the analyte and the internal standard are recorded Ratio of analyte signal/internal standard signal are plotted against the concentration of the analyte in the standard solutions

Ratio of analyte signal in the sample / internal standard is calculated

A linear equation (y=mx + b) is obtained

Concentration of the analyte is calculated by solving for x when y is the ratio of analyte signal/internal standard signal in the unknown sample

Either one standard or several standards, to create a calibration curve, may be used.

GC TECHNOLOGY: EXTERNAL STANDARD (ESTD)

An ESTD quantitation uses a known data from a calibration standard and unknown data from the sample are combined to generate a quantitative report.

It is called external standard because the standard material is external to the unknown material.



ESTD Response Factor Calculation:

 $\mathsf{RF} = (\mathsf{A}_{\mathsf{x}}) / (\mathsf{C}_{\mathsf{x}})$

 A_x = Area of the compound

 C_x = Concentration of the compound

SCD – External Standard

Back Signal Results					
Name	Retention Time	Area	ESTD concentration	CALC Resp Factor	
1propanethiol ,200,10	9.995	8177678	58.7 =	7.17808E-06	
nbutanethiol ,200,20	17.233	10399294	62.4	6.00041E-06	
2mthiophene ,200,22	20.559	8652322	52.1	6.0215E-06	
edisulfide ,200,37	27.492	18458636	55	2.97964E-06	
benzothiophene ,200,39	37.565	8705206	61.4	7.05325E-06	
light sulfur ,200,4		0	0		
heavy sulfur ,200,2		0	c 1		
Totals					
		54393136	289.6		
1\F7ChromData\Drainata\CC E1\[+\r00057100022000	u do'		
TEZCHIOMDala/Projects/GC 51/r	result/10995/100055800.151	1109907100000000	.07		
1\EZChromData\Projects\GC 51\F Back Signal Results	Result/169957100035804.151	1109957100055800	.02		
	Retention Time	Area	ESTD concentration	n-butanethiol CALC	1 propanethiol CAL
Back Signal Results				n-butanethiol CALC	1 propanethiol CAL
Back Signal Results Name	Retention Time	Area			
Back Signal Results Name ethanethiol ,200,7	Retention Time 4.541	Area 2954381	ESTD concentration	=	21.2
Back Signal Results Name ethanethiol ,200,7 thiophene ,200,19	Retention Time 4.541 14.393	Area 2954381 16890253	ESTD concentration 101.4	101.3	21.2 121.2
Back Signal Results Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22	Retention Time 4.541 14.393 20.558	Area 2954381 16890253 455228	ESTD concentration 101.4 2.7	101.3 2.7	21.2 121.2 3.3
Back Signal Results Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22 3mthiophene ,200,23 light sulfur ,200,4	Retention Time 4.541 14.393 20.558	Area 2954381 16890253 455228 234646	ESTD concentration 101.4 2.7 1.4	101.3 2.7	21.2 121.2 3.3
Back Signal Results Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22 3mthiophene ,200,23	Retention Time 4.541 14.393 20.558	Area 2954381 16890253 455228 234646 0	ESTD concentration 101.4 2.7 1.4 0	101.3 2.7	21.2 121.2 3.3
Back Signal Results Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22 3mthiophene ,200,23 light sulfur ,200,4	Retention Time 4.541 14.393 20.558	Area 2954381 16890253 455228 234646 0	ESTD concentration 101.4 2.7 1.4 0	101.3 2.7	121.2 3.3

SCD – External Standard

Back Signal Results					
Name	Retention Time	Area	ESTD concentration	CALC Resp Factor	
1propanethiol ,200,10	9.995	8177678	58.7	7.17808E-06	
nbutanethiol ,200,20	17.233	10399294	62.4	6.00041E-06	
2mthiophene ,200,22	20.559	8652322	52.1	6.0215E-06	
edisulfide ,200,37	27.492	18458636	55	2.97964E-06	
benzothiophene ,200,39	37.565	8705206	61.4	7.05325E-06	
light sulfur ,200,4		0	0		
heavy sulfur ,200,2		0	0		
Totals					
		54393136	289.6		
1\EZChromData\Projects\GC 51\F	Result\r899571c0033aoq.rsl	t\r899571c0033aoq	.dat		
Back Signal Results					
Buok Olghur Kooullo					
Name	Retention Time	Area	ESTD concentration	n-butanethiol CALC	1-propanethiol CAL
-	Retention Time 4.541	Area 2954381	ESTD concentration 17.7	n-butanethiol CALC 17.7	1-propanethiol CAL 21.2
Name					
Name ethanethiol ,200,7	4.541	2954381	17.7	17.7	21.2
Name ethanethiol ,200,7 thiophene ,200,19	4.541 14.393	2954381 16890253	17.7 101.4	17.7 101.3	21.2 121.2
Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22	4.541 14.393 20.558	2954381 16890253 455228	17.7 101.4 2.7	17.7 101.3 2.7	21.2 121.2 3.3
Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22 3mthiophene ,200,23	4.541 14.393 20.558	2954381 16890253 455228 234646	17.7 101.4 2.7 1.4	17.7 101.3 2.7	21.2 121.2 3.3
Name ethanethiol ,200,7 thiophene ,200,19 2mthiophene ,200,22 3mthiophene ,200,23 light sulfur ,200,4	4.541 14.393 20.558	2954381 16890253 455228 234646 0	17.7 101.4 2.7 1.4 0	17.7 101.3 2.7	121.2 3.3

SCD – External Standard

Back Signal Results					
Name	Retention Time	Area	ESTD concentration	CALC Resp Factor	
1propanethiol ,200,10	9.995	8177678	58.7	7.17808E-06	
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edisulfide ,200,37	27.492	<mark>18458636</mark>	<mark>55</mark>	2.97964E-06	
benzothiophene ,200,39	37.565	8705206	61.4	7.05325E-06	
light sulfur ,200,4		0	0		
heavy sulfur ,200,2		0	0		
Totals					
		54393136	289.6		
1\EZChromData\Projects\GC 51\I	Result/r899571c0033aoq.rsl	t\r899571c0033aoq	.dat		
Back Signal Results					
Name	Retention Time	Area	ESTD concentration	n-butanethiol CALC	1-propanethiol CAL
ethanethiol ,200,7	4.541	2954381	17.7	17.7	21.2
	14.393	16890253	101.4	101.3	121.2
thiophene ,200,19	14.000				
thiophene ,200,19 2mthiophene ,200,22	20.558	455228	2.7	2.7	3.3
· ·			2.7 1.4	2.7 1.4	3.3 1.7
2mthiophene ,200,22	20.558	455228			
2mthiophene ,200,22 3mthiophene ,200,23	20.558	455228 234646	1.4		
2mthiophene ,200,22 3mthiophene ,200,23 light sulfur ,200,4	20.558	455228 234646 0	1.4 0		
2mthiophene ,200,22 3mthiophene ,200,23 light sulfur ,200,4	20.558	455228 234646 0	1.4 0		

GC TECHNOLOGY: NORMALIZATION

Different components have different response factors to compensates for different detector response for different components. These factors may be calculated by preparing a synthetic mixture or by calculating a theoretical response factor.



Normalization Calculation:

$$\% X = \frac{Ax.fx}{\Sigma Af} x \ 100$$

 $\frac{Ax \text{ is Area . } fx \text{ is response factor}}{\Sigma Af \text{ is Sum of all RF modified areas}} x 100$

FID - Normalization

\\ims01\EZChromData\Projects\GC 46\Result\r898280c0039udk.rslt\r898280c0039udk.dat

Back Signal Results					
Name	Retention Time	Area	RRF	NORM Area	NORM concentration
ethene ,100,28	13.233	197236	0.966		
		24383	1		
propane ,100,34	15.875	4542	1.01		
isobutane ,100,48	20.313	146189	1		
1butene ,100,40	23.303	10481868	0.966		
		8852	1		
nbutane ,100,49	24.515	122391584	1		
		16738	1		
t2butene ,100,45	25.868	155159516	0.966		
		226016	1		
22dmpropane ,100,69	26.562	21733	0.994		
c2butene ,100,42	27.484	69035406	0.966		
isopentane ,100,71	32.432	4068	0.994		
2m2butene ,100,62	36.172	13229	0.966		
3m1pentene ,100,63	38.591	5458209	0.965		
		41	1		
23dm-butane ,100,106	39.045	6435	0.994		
2m-pentane ,100,107	39.279	35130	0.994		
3m-pentane ,100,93	40.034	99455	0.994		
1-hexene ,100,84	40.317	1830098	0.966		
n-hexane ,100,109	40.823	16327607	0.966		
		6327414	1		
t-3-hexene ,100,102	40.937	8337254	0.966		
c-3-hexene ,100,95	41.005	1219428	0.966		
t-2-hexene ,100,101	41.065	17768128	0.966		
				405415994	100.0000

FID - Normalization

\\ims01\EZChromData\Projects\GC 46\Result\r898280c0039udk.rslt\r898280c0039udk.dat

Back Signal Results					
Name	Retention Time	Area	RRF	NORM Area	NORM concentration
ethene ,100,28	13.233	197236	0.966	190529.976	
		24383	1	24383	
propane ,100,34	15.875	4542	1.01	4587.42	
isobutane ,100,48	20.313	146189	1	146189	
1butene ,100,40	23.303	10481868	0.966	10125484.49	
		8852	1	8852	
nbutane ,100,49	24.515	122391584	1	122391584	
		16738	1	16738	
t2butene ,100,45	25.868	155159516	0.966	149884092.5	
		226016	1	226016	
22dmpropane,100,69	26.562	21733	0.994	21602.602	
c2butene ,100,42	27.484	69035406	0.966	66688202.2	
isopentane ,100,71	32.432	4068	0.994	4043.592	
2m2butene ,100,62	36.172	13229	0.966	12779.214	
3m1pentene ,100,63	38.591	5458209	0.965	5267171.685	
		41	1	41	
23dm-butane ,100,106	39.045	6435	0.994	6396.39	
2m-pentane ,100,107	39.279	35130	0.994	34919.22	
3m-pentane ,100,93	40.034	99455	0.994	98858.27	
1-hexene ,100,84	40.317	1830098	0.966	1767874.668	
n-hexane ,100,109	40.823	16327607	0.966	15772468.36	
		6327414	1	6327414	
t-3-hexene ,100,102	40.937	8337254	0.966	8053787.364	
c-3-hexene ,100,95	41.005	1219428	0.966	1177967.448	
t-2-hexene ,100,101	41.065	17768128	0.966	17164011.65	
					_

FID - Normalization

\\lims01\EZChromData\Projects\GC 46\Result\r898280c0039udk.rslt\r898280c0039udk.dat

Back Signal Results					
Name	Retention Time	Area	RRF	NORM Area	NORM concentration
ethene ,100,28	13.233	197236	0.966	190529.976	0.0470
		24383	1	24383	0.0060
propane ,100,34	15.875	4542	1.01	4587.42	0.0011
isobutane ,100,48	20.313	146189	1	146189	0.0361
1butene ,100,40	23.303	10481868	0.966	10125484.49	2.4976
		8852	1	8852	0.0022
nbutane ,100,49	24.515	122391584	1	122391584	30.1891
		16738	1	16738	0.0041
t2butene ,100,45	25.868	155159516	0.966	149884092.5	36.9704
		226016	1	226016	0.0557
22dmpropane ,100,69	26.562	21733	0.994	21602.602	0.0053
c2butene ,100,42	27.484	69035406	0.966	66688202.2	16.4493
isopentane ,100,71	32.432	4068	0.994	4043.592	0.0010
2m2butene ,100,62	36.172	13229	0.966	12779.214	0.0032
3m1pentene ,100,63	38.591	5458209	0.965	5267171.685	1.2992
		41	1	41	0.0000
23dm-butane ,100,106	39.045	6435	0.994	6396.39	0.0016
2m-pentane ,100,107	39.279	35130	0.994	34919.22	0.0086
3m-pentane ,100,93	40.034	99455	0.994	98858.27	0.0244
1-hexene ,100,84	40.317	1830098	0.966	1767874.668	0.4361
n-hexane ,100,109	40.823	16327607	0.966	15772468.36	3.8904
		6327414	1	6327414	1.5607
t-3-hexene ,100,102	40.937	8337254	0.966	8053787.364	1.9865
c-3-hexene ,100,95	41.005	1219428	0.966	1177967.448	0.2906
t-2-hexene ,100,101	41.065	17768128	0.966	17164011.65	4.2337

BASIC GC EVENT FLOW

Sample Introduction

Liquid/Gas Valve

Syringe

Manual/Automatic

Separation Process

Capillary Column

Packed Column

Multi-Valve/Column

Component Detection

Flame "FID"

Thermal "TCD"

Sulfur "SCD", "FPD"

Many Others

Data Handling References

Calibration

Quantification

TITRATION

Characterization of single component in mixtures

Water Petroleum liquids Many others

Applications cover analysis of

Liquid streams Water analysis Environment samples Final Product



Overview – What is A Titration?

What is a titration?

- A technique where a solution of known concentration is used to determine the concentration of an unknown solution.
- Typically, the titrant (the known solution) is added from a burette to a known quantity of the analyte (the unknown solution) until the reaction is complete.
- Knowing the volume of titrant used allows the determination of the unknown concentration.
- Often, an indicator is used to signal the end of the reaction, the endpoint.
- An automatic titrator uses potentiometric changes to determine the endpoint.

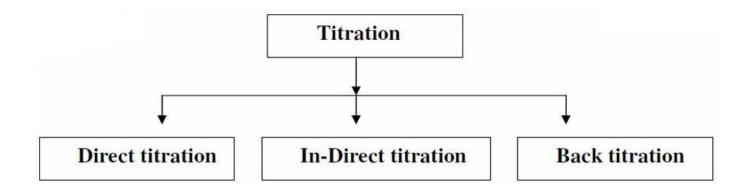
Example: Determination of acetic acid (CH₃COOH) by titration with sodium hydroxide (NaOH)

$$\begin{array}{cccc} CH_{3}COOH + NaOH \\ (analyte) & (titrant) \end{array} \xrightarrow{} \begin{array}{c} CH_{3}COONa + H_{2}C \\ (reaction products) \end{array}$$

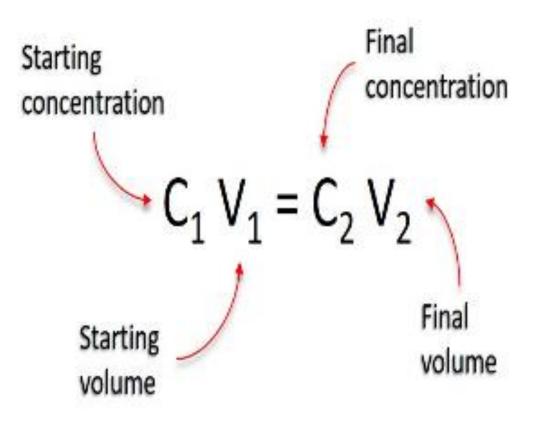
Overview – Methods of Titration

•Methods of Titration:

•There are of three types of titration based on the method used in the process of titration.



Overview – Titration Types



- Direct Titration:
 - A conventional titration where a known amount of titrant is added from a burette to a sample taken in a flask.
 - One substance is analyzed for its quantity by another substance of known volume and concentration.
- Indirect Titration:
 - Theoretically, it is converting a substance into acid and analyzing it with a base. (vice-versa).
 - This is a method for non readily reactive substances.
 - If a substance is weakly acidic it might not permit a precise analysis by direct titration.
 - So first that substance is chemically altered to be more reactive in acidic or basic form and then analyzed by adding a titrant.
- Back Titration:
 - This method is suitable for weakly reactive or non-reactive substances.
 - The sample is allowed to react with excess and known quantity of a titrant before starting the titration.
 - The remaining excess base or acid is estimated by a known quantity of acid or base back titrated to an endpoint.

Overview -Titration Types

Volumetric Titration:

• With volumetric titration, the titrant is added to the sample by a burette from an external source. The volume of titrant added to the sample is measured during the titration.

Coulometric Titration:

 In coulometric titration, the titrant is generated electrochemically in the sample within in the titration cell. This means that a precursor of the titrant that reacts with the analyte is already present in the sample before the analysis starts.

Overview – Titration Types

• Advantages of Titration:

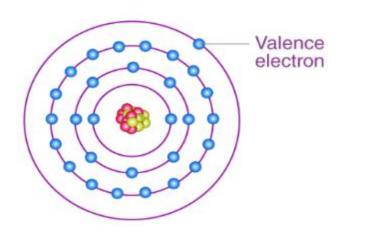
- There are several reasons why titration is used in laboratories worldwide:
 - Titration is an established analytical technique.
 - It is fast.
 - It is a very accurate and precise technique.
 - A high degree of automation can be implemented.
 - Titration offers a good price/performance ratio compared to more sophisticated techniques.
 - It can be used by low-skilled and low-trained operators.
 - No need for highly specialized chemical knowledge.

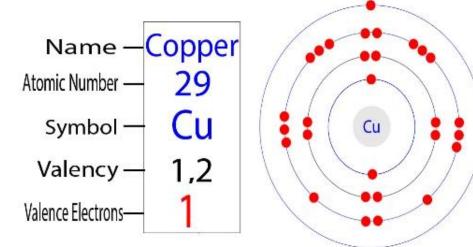
- Buffer Solution:
 - A solution that resists changes in pH even when a strong acid or base is added or when it is diluted with water.
- Analyte:
 - The chemical substance within our test sample to be determined with the analysis.
 - Example: Mercaptan sulfur
- Matrix:
 - Everything in the sample besides the analyte(s). Side reactions can occur due to other components.
 - Matrix Examples: Water, hydrocarbons, acids, etc.
- Titrant:
 - Solution of known concentration which is added to the sample (titration) and reacts with the analyte. The analyte content is calculated from the consumption of the titrant.
 - Titrant Examples: KOH, BrBrO3, AgNO3, etc.



• Valence Electrons:

- Valence electrons are electrons located in the outermost electron shell of an atom.
- These electrons, being the furthest from the nucleus and thus the least tightly held by the atom, are the electrons that participate in bonds and reactions.
- The number of valence electrons that an element has determines its reactivity, electronegativity, and the number of bonds it can form.

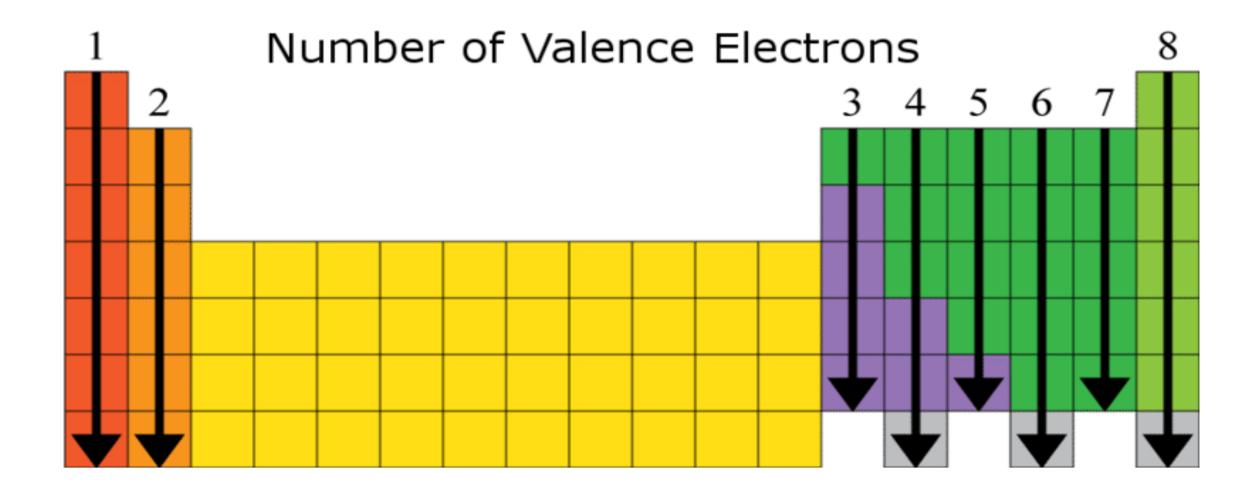




Valence Electrons Continued:

	1 H Hydrogen 1	2		Atomi No.	c	→ 1	н ←		Symb	01			13	14	15	16	17	18 2 He natur 2
	Li Litum	4 Be Beryllium 2		Name	·	> Hyd	drogen 1 ←		Valer	ice El	ectro	n	B Boren 3	A C Carbon 4	7 Nitropen 5	O Convert 6	F Flaorine 7	Ne Ne
	Na Sedum	12 Mg Magnetium 2	3	4	5	6	7	8	9	10	11	12	Al Aluminium 3	Si Silocan	LS P Phosphorus 5	Solphat 6	Cl Chiarine 7	Ar Argon
	K Potassium 1	20 Ca Cakture 2	Scardum	Ti Titanium 4	Z3 V Vanachum	Cr Chromham 6	25 Mn Mangariese	Fe ton 8	27 Co Cobalt	Ni Nichel 10	Cu Copper 11	300 Zn 2014 12	Ga Gatum	Ge Germantum 4	AS Arsenic 5	Selanium 6	Br Bromine 7	In Kr Krypto 8
	Rb Rubictium	38 Sr Strooman	30 ¥ Yttmam 3	40 Zr Zircontum 4	HI ND Nobium	42 Mo Molyischen	43 TC Technedum	Ru	45 Rh Rhodium 9	Pd Paladum 10	47 Ag Silver 11	48 Cd Cadmium 12	47 In Indium	50 50 70 4	SI Sb Antimony 5	52 Te Tellurium 6	5) I Jodine 7	S4 Xe Xenor 8
	ST CS Carestam	Se Ba Bartum Z	57 La Lanthanum 3	Hf	Ta Tantalum 5	74 W Tungstee	To Re Rhosture 7	TO Os Osmium 8	77 Ir Indum 9	Pt Pt Platnum 10	74 Au 00M 11	Hg Mercury 12	TI Thallium	Pb Lead	83 Bi Bismath 5	PO Pokorium 6	At Astartine 7	Rn Rador 8
	Fr	ti Ra Radium	Actinum	104 Rf Ratherfor	Dubcium	105 Sg	Bh Botelam	Haserium	109 Mt Motneriam	Ds Dermitad	Rg	Cn Copermici	Nhprium	Fierceitem	Mc Motosylum	LV Lv	TS Tennessine	Ogarress
1	Lanti	hanid	des	4 58 Ce Cerium 4	5 Pr Prinseody 5	6 Nd Neodymi. 6	61 Pm Promethi. 7	8 Sm Samarium 8	e3 Eu	10 Gd Oadolmin 10	11 Tb Sentsum 11	12 Dy Dytercolum 12	a7 Ho Hothum	4 Er Er 14	5 Tm Thuliam 15	70 Yb Yberbium 16	71 Lu Lutetum 3	8
	Act	inide	25	90 Th Thorium	PI Pa Protactini.	92 U Liranium	93 Np Neptunium 7	94 Pu Plutonium 8	95 Am Americum 9	Cm Curium 10	97 Bk Berkelum 11	os Cf Californium 12	95 ES Einsteinium 13	Fermium 14	Md Mendeley 15	No Notelium 16	Lr Lawrenc	

Valence Electrons Continued:





Molarity:

- The number of moles of solute per liter of solution.
- The units of molarity are M or mol/L. A 1 M solution is said to be "one molar."

Normality:

- The number of grams equivalent to solute that is present in a one-liter solution.
- Common units of normality include N, eq/L, or meq/L.

Molarity to Normality Conversions:

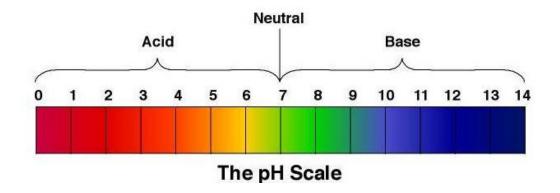
- When doing normality and molarity conversions, remember when the valence is one, the normality and molarity are the same. Thus, for NaOH, it is the same.
 - Normality = Valence x Molarity
 - Molarity = Normality/Valence

- Molarity/Normality Conversions Examples:
 - NaOH Example:
 - To go from 0.1 molarity to normality:
 - valence = 1
 - Molarity = 0.1
 - Therefore (1 valence) x (0.1 molarity) = 0.1 Normality.
 - To go from 0.1 <u>normality to molarity:</u>
 - valence = 1
 - Molarity = 0.1
 - Therefore (0.1 normality) / (1 valence) = 0.1 Molarity

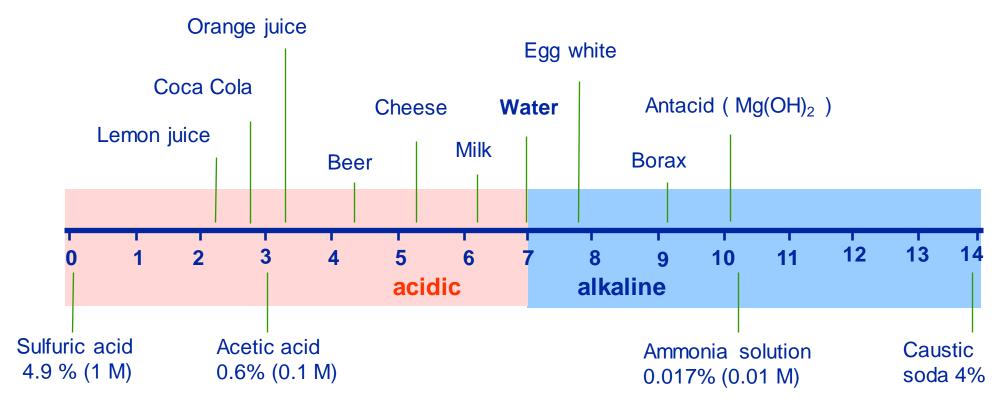
- Molarity Normality Conversions Examples Continued:
 - H₂SO₄ Example:
 - To go from 0.1 molarity to normality:
 - valence = 2
 - Molarity = 0.1
 - Therefore (2 valence) x (0.1 molarity) = 0.2 Normality.
 - To go from 0.1 <u>normality to molarity:</u>
 - valence = 2
 - Normality = 0.1
 - Therefore (0.1 normality) / (2 valence) = 0.05 Molarity

- **pH:** The negative logarithm of H+ ion concentration. (pH = -log [H⁺])
- pH Scale:
- Hydronium Ion vs Hydroxide Ion
 - For acidic solution: $[H_3O^+] > [OH^-]$
 - For neutral solution: $[H_3O^+] = [OH^-]$
 - For basic solution:

 $[H_3O^+] > [OH^-]$ $[H_3O^+] = [OH^-]$ $[H_3O^+] < [OH^-]$



• pH of Typical Solutions:

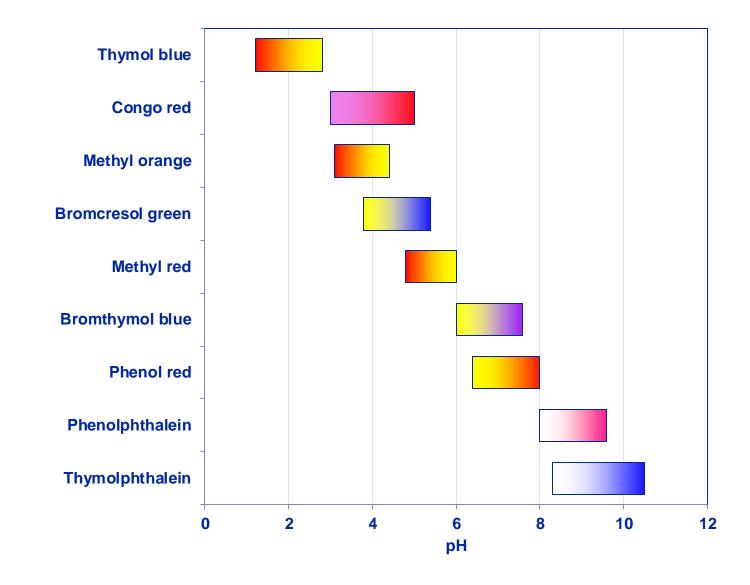




- Indications of Inflection Point:
 - The equivalence point of a titration should be easily visible or measurable.
 - The two basic indication principles are:
 - Indicator:
 - An indicator is a substance which changes its color at the equivalence point.
 - Small amounts of an indicator are added to the solution prior to the analysis.
 - Example:
 - Phenolphthalein is an indicator which is pink under basic conditions and colorless under acidic conditions.
 - At the equivalence point of an acetic acid titration by sodium hydroxide, the color will immediately change from colorless to pink.

• pH Indicators:

For a manual titrations, various pH indicators exist. The selection must be based on the pH range where the equivalence point occurs. The indicator must be chosen such that its color change occurs at the equivalence point. The following graph shows common pH indicators and their color change range:



Indications of Inflection Point Continued:

Electrode: (potentiometric change)

An electrode is an instrument that measures a specific property of a solution electrochemically.

A suitable electrode must be chosen for each type of reaction.

The electrode must measure a property which is related to the titration reaction.

The whole titration procedure can be followed with an electrode by a titration curve

Example:

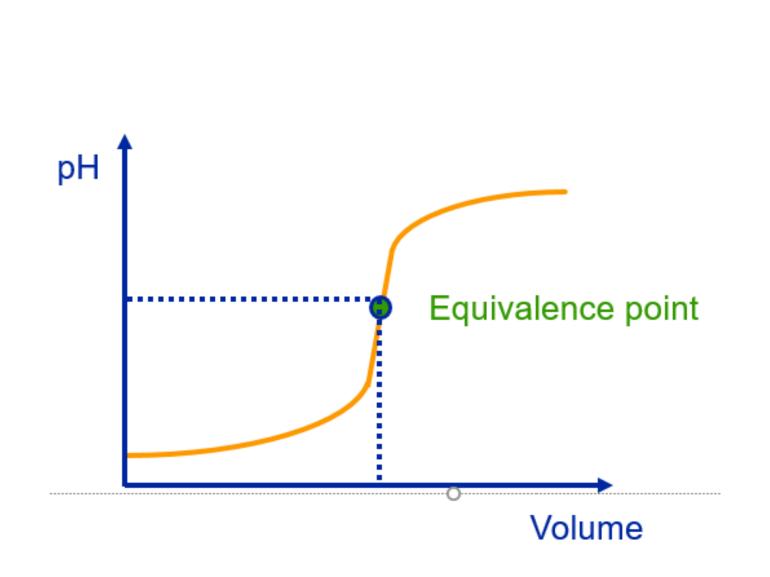
For the titration of acetic acid by sodium hydroxide a pH electrode is used. The pH electrode measures the H+ concentration related to acetic acid that hasn't reacted. The equivalence point is reached when the pH value changes suddenly from acidic to basic.



Indications of Inflection Point

Continued:

- If an electrode is used for a titration, the signal vs. titrant volume can be plotted.
- Such a plot is called a titration curve.
- The equivalence point can be determined directly from a titration curve and is the inflection point of the curve.
 - Example: Titration curve (pH vs. volume) of acetic acid titration with sodium hydroxide. The equivalence point is clearly visible.



Two different titration terminations are used:

- Equivalence Point Titration (EQP)
- Endpoint Titration (EP)

EQP:

- The EQP is reached as soon as all analyte has reacted with the titrant.
- In a titration curve, the EQP is the inflection point of the curve.
- The titration is carried out over the EQP and evaluated afterwards.

EP:

- The EP is reached as soon as the signal reaches a predefined value.
- The titration is usually stopped at the EP.

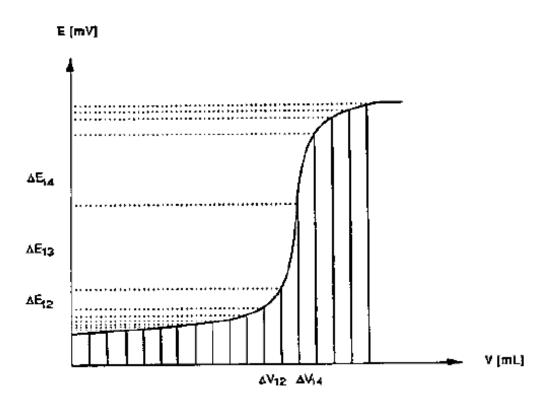
• Equivalence Point (Inflection Point):

- The point when enough reagent is added to <u>react</u> <u>completely</u> with the analyte.
- At the equivalence point, the number of moles of the titrant are equal to the number of moles of analyte.
- It occurs before the endpoint.
- It does not indicate the completion of a titration process.
- Equivalence point means the completion of reaction between titrant and analyte.
- A titration process can have multiple equivalence points.
- Just before change in color.
- For calculation of the result the equivalent number z has to be known.
- The equivalent number of an analyte is determined by the stoichiometry. In general, the equivalent number is the number of moles "monovalent" titrant (e.g., NaOH, HCI, AgNO₃, Na₂S₂O₃) which are needed for the reaction with one mole of analyte.

•Equivalence Point (Inflection Point)

INFLECTION

INCREMENT: CONSTANT SIZE LARGE

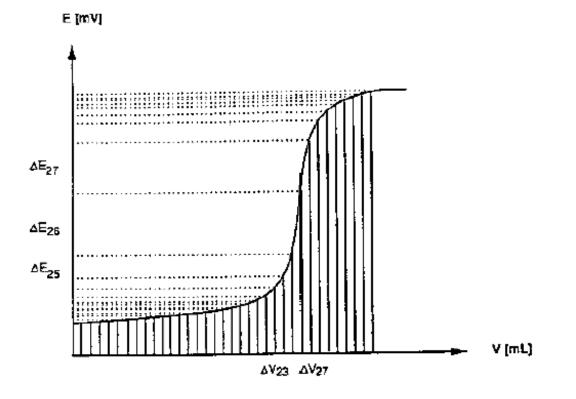


_∆V = constant

•Equivalence Point (Inflection Point)

INFLECTION

INCREMENT: CONSTANT SIZE SMALL

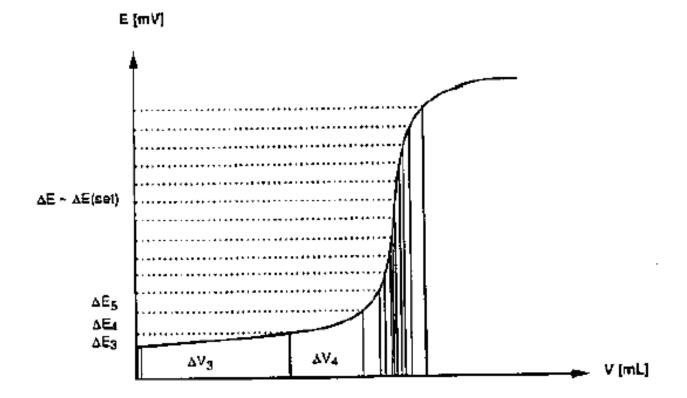




•Equivalence Point (Inflection Point)

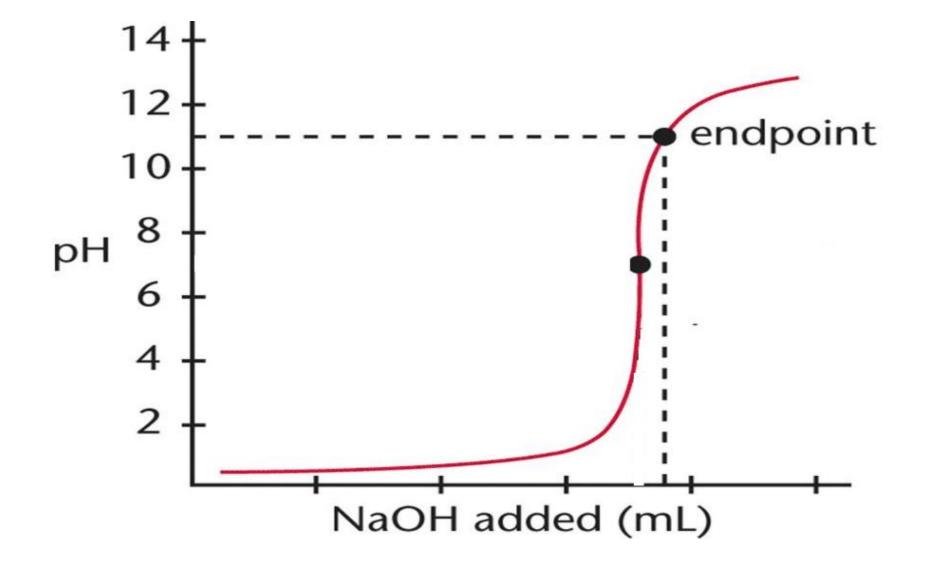
INFLECTION

INCREMENT: DYNAMIC VARIABLE



• End point:

- The point at which color change i.e. (medium change) occurs in the system due to pH change is called endpoint
- At this point, the moles of titrant <u>exceed</u> the moles of the analyte. A sharp change in pH occurs resulting in color change.
- It comes after equivalence point.
- Titration is complete once the endpoint is reached.
- Endpoint does not mean the completion of reaction between analyte and titrant.
- It occurs once in a reaction.
- Change in color.



Categories	Endpoint	Equivalence point			
Definition	The point at which color change occurs due to a pH- change	A point when the chemical reaction in the titration mixture ends			
Explanation	At this point, the moles of titrant exceed the moles of the analyte.	A point in a titration when the number of moles of titrant are equal to analyte			
Occurrence	After equivalence point	Before the endpoint			
Completion	Titration is complete once the endpoint reaches	It is not the completion of a titration reaction			
Reaction completion	It does not mean the completion of reaction between analyte and titrant	It means the completion of reaction between titrant and analyte			
Times of occurrence	Occurs only once in a reaction	Titration processes can have multiple equivalence points			
Nutshell	Change in color	Just before change in color			

Titration Reactions Need To Be:

Selective:	•The chemical reaction between the analyte and the titrant must be selective (only the analyte should react with the titrant).
Fast:	•The reaction should be fast in order to guarantee that the added titrant reacts immediately with the analyte.
Complete:	•The equilibrium of the reaction should lie strongly on the product side of the reaction to guarantee a complete reaction.
Unambiguous:	•The stoichiometry of the reaction must be known and unambiguous.
Accurate:	 The key point of a good titration is the accurate determination of the titrant volume used. For this reason, the following two requirements must be fulfilled: A titrant addition in small quantities must be possible. An accurate reading for the volume used is needed.

Acid-Base Titration Reaction

(Neutralization Reaction):

What is an Acid-Base Titration?

A quantitative analysis for determining the concentration of an acid or base by neutralizing it with a standard solution of base or acid having known concentration according to a stoichiometric proton-transfer reaction.

Acid-base reactions involve the transfer of hydrogen ions between reactants.

ACID/BASE REACTIONS

$HCl + NaOH \longrightarrow NaCl + H_2O$

ACID + BASE → SALT + WATER

- Oxidation-Reduction Reaction (Redox): What is a Redox Reaction (Oxidation-Reduction)?
 - A Redox reaction is a chemical reaction that involves transfer of electrons between two species.
 - The oxidation number of a molecule, atom, or ion changes by gaining or losing an electron.
 - Redox reactions involve a change in oxidation number for one or more reactant elements.

Reduction H_2 Fe

Oxidation

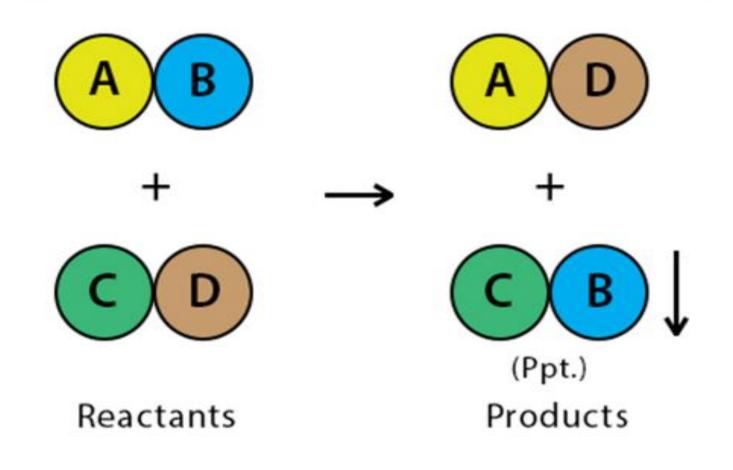
• Precipitation Reaction:

What is a Precipitation Reaction?

- Reactions that involve the formation of one or more insoluble products.
- Reactions that occur when cations and anions combine to form an insoluble ionic solid called a precipitate.
- Many reactions of this type involve the exchange of ions between ionic compounds in aqueous solution and are sometimes referred to as double displacement, double replacement, or metathesis reactions.

Example: Removing H2S for Mercaptan titration.

Precipitation Reaction



• Examples of

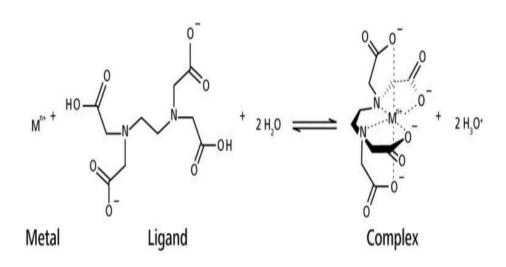
Precipitation Reactions:

Double-replacement Reaction Examples

KBr +	AgNO ₃ →	KNO ₃	+ AgBr↓
Potasium	Silver	Potassium	Silver
bromide	nitrate	nitrate	bromide
Na ₂ SO ₄ + Sodium sulfate	Strontium chloride	2 NaCl Sodium chloride	+ SrSO₄↓ Strontium sulfate
HCI + Hydrochloric acid	NaOH — Sodium hydroxide	NaCl Sodium chloride	+ H ₂ O Water
Na2S +	2 HCI →	2 NaCl	+ H₂S↑
Sodium	Hydrochloric	Sodium	Hydrogen
sulfide	acid	chloride	sulfide

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- Complexometric Reaction:
 - What is a Complexometric reaction?
 - When a simple ion is transformed into a complex ion and the equivalence point determined by using metal indicators or electrometrically.



Complexometric reactions: $Mg^{2+} + EDTA \rightarrow Mg[EDTA]^{2+}$ $Ca^{2+} + EDTA \leftrightarrow Ca[EDTA]^{2+}$ Examples: Total hardness of water (Mg²⁺ and Ca²⁺) Calcium content in milk and cheese Cement analysis

•
$$R = \frac{VEQ \cdot c \cdot t \cdot C}{m}$$

For Direct Titration Calculations:

- The analyte content in a direct titration is calculated from the titrant consumption at the equivalence or end point using the following formula:
 - = Result R
 - *VEQ* = Titrant consumption at the equivalence or end point (in mL)
 - = Titrant concentration (in mol/L) С t
 - = Titrant titer (no unit)
 - = Constant for unit conversion С
 - = Sample size (in g or mL) m
- The constant C is dependent on the result unit wanted and on the unit of the sample size

Calculations

- Examples of Constants:
- M = Molar mass of the analyte in g/mol
- z = Equivalent number (no unit)

Sample size entry unit	g	mL	
	%		$C = \frac{M}{10 \cdot z}$
ij	mg/g	g/L	$C = \frac{M}{z}$
Result unit	ppm	mg/L	$C = \frac{M \cdot 1000}{z}$
Re	mmol/kg	mmol/L	$C = \frac{1000}{z}$
	mol/kg	mol/L	$C = \frac{1}{z}$

Calculations

- Example 1:
 - For the titration of the acetic acid (M = 60.04 g/mol) in a 1 g sample of vinegar, 5 mL sodium hydroxide (c = 0.1 mol/L, titer: 0.9900) were consumed until the equivalence point was reached.
 - To calculate the acetic acid content in % you have to use the following formula:

$$C = \frac{M}{10 \cdot z} = \frac{60.04 \text{ g/mol}}{10 \frac{\text{mg}}{\text{g} \cdot \%} \cdot 1} = 6.004 \frac{\text{g} \cdot \%}{\text{mmol}}$$

$$R = \frac{VEQ \cdot c \cdot t \cdot C}{m} = \frac{5 \text{ mL} \cdot 0.1 \frac{\text{mol}}{\text{L}} \cdot 0.9900 \cdot 6.004 \frac{\text{g} \cdot \%}{\text{mmol}}}{1 \text{ g}} = 2.97 \%$$

Calculations

• Titer Determinations:

- The **titer** is usually determined by a direct titration, but the calculation is a little different.
- The titer (*t*) is defined as the actual concentration (c_{act}) divided by the nominal concentration (c_{nom}) of a titrant:
- The following formula can be used to calculate the titer of a titrant, if this titrant was used to titrate a titer standard by a direct titration:
 - t = Titrant titer (no unit)
 - *m* = Titer standard sample size (in g or mL)
 - *VEQ* = Titrant consumption at the equivalence or end point (in mL)
 - *c* = (Nominal) titrant concentration (in mol/L)
 - *C* = Constant for unit conversion

 $t = \frac{c_{\rm act}}{c_{\rm nom}}$

 $t = \frac{m}{VEQ \cdot c \cdot C}$

• Titer Determinations Continued:

• If the titer standard substance is a solid, use the following formula for the constant C:

$$C = \frac{M}{10 \cdot p \cdot z}$$

• For liquid titer standards the following constant is used:

$$C = \frac{1}{cst \cdot z}$$

- M = Molar mass of the analyte (in g/mol)
- p = Purity of the solid titer standard (in %)
- z = Equivalent number of the titer standard (no unit)
- *cst* = Concentration of the liquid titer standard (in mol/L)

• Example 2:

R

- Potassium hydrogen phthalate (KHP, M= 204.23 g/mol) was used as a solid titer standard to determine the titer of sodium hydroxide (c = 0.1 mol/L). 0.0931 g of KHP (purity: 99 %) was weighed into a titration beaker and diluted with deionized water.
- This solution was titrated with sodium hydroxide. The equivalence point was detected at a titrant consumption of 4.5238 mL. For this measurement the titer can be calculated as follows:

$$C = \frac{M}{10 \cdot p \cdot z} = \frac{204.23 \frac{g}{mol}}{10 \frac{mg}{g \cdot \%} \cdot 99 \% \cdot 1} = 0.206 \frac{g}{mmol}$$
$$= \frac{m}{VEQ \cdot c \cdot C} = \frac{0.0931 g}{4.5238 \text{ mL} \cdot 0.1 \frac{mol}{L} \cdot 0.206 \frac{g}{mmol}} = 0.9990$$

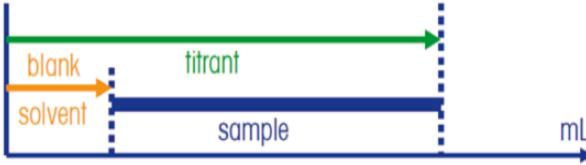
Calculations - Blanks

In some titrations it may happen that the solvent itself also reacts with the titrant. The amount of titrant used for the solvent is called the **blank value**.

The blank value must be **compensated to get the correct result**.

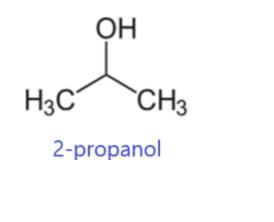
To determine the blank value a titration of the **solvent without any sample** has to be performed. The blank value is the volume of the titrant used until the equivalence or endpoint is reached.

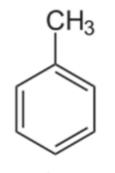
For a titration where a blank value is used, take care to always use the same solvent volume!



Calculations - Blanks

- Example 3:
 - The acid number of motor oil is determined by a non-aqueous acid/base titration with potassium hydroxide (in 2propanol) as titrant.
 - The solvent mixture of toluene, 2-propanol and water is used.
 - Before titrating the sample, the solvent mixture without any sample is titrated.
 - The titrant consumption of this blank measurement is compensated in the calculation of the sample measurement.





Toluene



Calculations - Blanks

• The calculation of a blank value compensated titration is similar to the direct titration:

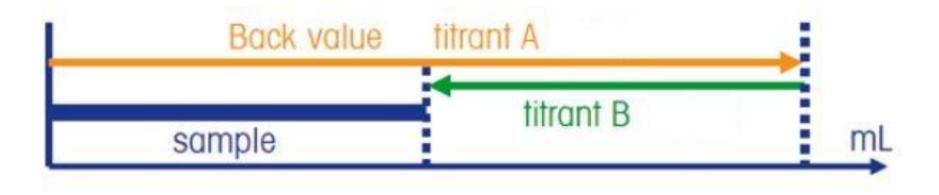
$$R = \frac{(VEQ - B) \cdot c \cdot t \cdot C}{m}$$

R = Result

- VEQ = Used titrant volume until the equivalence or end point (in mL)
- B = Blank value, titrant volume used for the titration of the solvent (in mL)
- *c* = Titrant concentration (in mol/L)
- t = Titrant titer (no unit)
- *C* = Constant for unit conversion
- *m* = Sample size (in g or mL)
- The constant *C* is the same as for the direct titration

Calculations – Back Titrations

- In a back titration a known excess amount of reagent (titrant A with known concentration) is added to the sample. This titrant reacts with the analyte. Afterwards the amount of non-reacted titrant A will be titrated with a second titrant (titrant B).
- The added volume of titrant A must be known very precisely.
- For accurate measurements, a so called **back value** is determined by titrating the added volume of titrant A without any sample. The result is based on the used volume of titrant B without any sample.
- Such titrations are often used when the reaction between the analyte and the first titrant is slow.



Calculations – Back Titrations

- Example 4:
 - For the titration of nitrite in soya sauce a known volume of potassium permanganate (titrant A) is added to the sample.
 - Potassium permanganate will react with the nitrite and the excess of the permanganate is titrated with ammonium ferrous sulfate(II) (titrant B).

$$5 \text{ NO}_2^- + 2 \text{ MnO}_4^- + 6 \text{ H}^+ 2 \text{ Mn}^{2+} + 5 \text{ NO}_3^- + 3 \text{ H}_2\text{O}$$

(analyte) (titrant A)

 $MnO_{4^{-}} + 5 Fe^{2+} + 8 H^{+} \longrightarrow Mn^{2+} + 5 Fe^{3+} + 4 H_2O$ (titrant A) (titrant B)



Calculations – Back Titrations

• The analyte content in a back titration is calculated using the following formula:

$$R = \frac{(Bk - VEQ) \cdot c \cdot t \cdot C}{p \cdot m}$$

R = Result

Bk = Back value, volume of titrant B used for the titration of titrant A without sample (in mL)

- VEQ = Volume of titrant B used to back titrate the excess of titrant A after sample addition (in mL)
- c = Concentration of titrant B (in mol/L)
- t = Titer of titrant B (no unit)
- *C* = Constant for unit conversion (unit part, unit is depending on the result needed)
- *p* = Stoichiometric proportion between titrant A and titrant B of the second reaction, stoichiometric factor of titrant B divided by factor of titrant A (no unit).
- *m* = Sample size (in g or mL)
- The constant C is the same as for the direct titration

- For calculation of the result the equivalent number, z must be known.
- The equivalent number of an analyte is determined by the stoichiometry. In general, the equivalent number is the number of moles "monovalent" titrant (e.g., NaOH, HCI, AgNO3, Na2S2O3) which are needed for the reaction with one mole of analyte.
- For acid/base titrations the equivalent number is the number of protons (H+) an acid releases by complete dissociation or the number of protons a base gains by reaction with an acid.
- For redox titrations the equivalent number is given by the number of electrons which are released or gained by a reductant or oxidant.
 - Example 1: Sulfuric acid is a diprotic acid.

 $H_2SO_4 + 2 NaOH$ \swarrow $Na_2SO_4 + 2 H_2O$ z = 2

• Example 2: For the reduction of Cr6+ to Cr3+ three electrons are needed.

 $Cr^{6+} + 3e^- \longrightarrow Cr^{3+} \qquad z = 3$

- If a titrant is "polyvalent" the equivalent concentration (normality) must be used for calculation.
- The equivalent concentration of titrant A is calculated by the following formula, where z is the equivalent number of the titrant:

 $c(1/z \mathbf{A}) = c(\mathbf{A}) \cdot z$

- Example 1: Sulfuric acid (c(H2SO4) = 0.05 mol/L) as titrant:

 $H_2SO_4 \longrightarrow SO_4^{2-} + 2 H^+$

$$c(1/2 \text{ H}_2\text{SO}_4) = c(\text{H}_2\text{SO}_4) \cdot z = 0.05 \frac{\text{mol}}{\text{L}} \cdot 2 = 0.1 \frac{\text{mol}}{\text{L}}$$

- Example 2: Potassium permanganate (c(KMnO4) = 0.02 mol/L) as titrant:

$$KMnO_4 + 8 H^+ + 5 e^- \longrightarrow Mn^{2+} + 4 H_2O + K^+$$

$$c(1/5 \text{ KMnO}_4) = c(\text{KMnO}_4) \cdot z = 0.02 \frac{\text{mol}}{\text{L}} \cdot 5 = 0.1 \frac{\text{mol}}{\text{L}}$$

- To calculate the content of the analyte, the titrant concentration must be known very accurately.
- To determine the exact titrant concentration, a titer determination is carried out prior to the sample analysis.
- For the titer determination, a substance of high purity (titrimetric standard) is titrated.
 - Example: The titer of sodium hydroxide is determined by titrating potassium hydrogen phthalate.

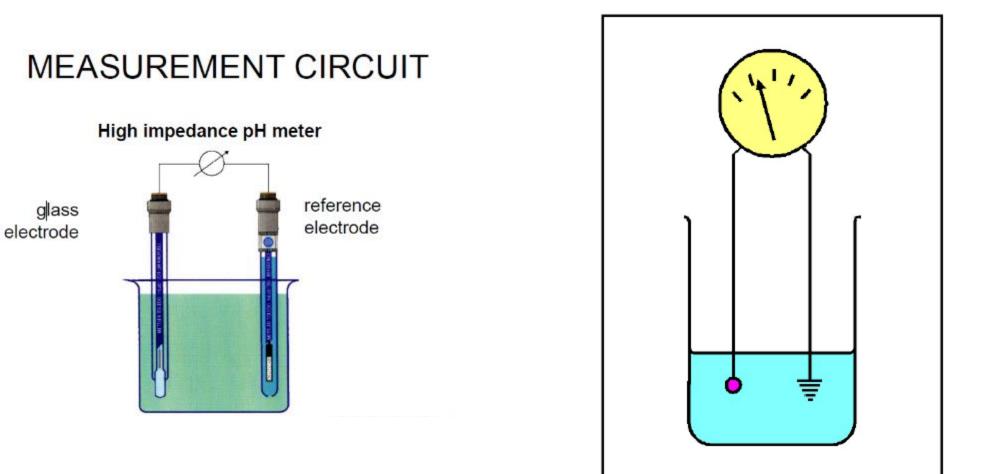
 $KOOC-C_6H_4-COOH + NaOH \longrightarrow KOOC-C_6H_4-COONa + H_2O$

• The actual titrant concentration (cact) is the nominal concentration (cnom) times the titer (t)

 $c_{\rm act} = c_{\rm nom} \cdot t$

Probes

- Probes (Electrodes):
 - Probes basically measure the current change between the analyte and the reference.



Probes – Reference vs Indicator

- What is the difference between Reference Electrodes and Indicator Electrodes?
 - Indicator electrode responds to changes in the activity of the analyte.
 - Reference electrode does not respond to changes, and its response is stable.





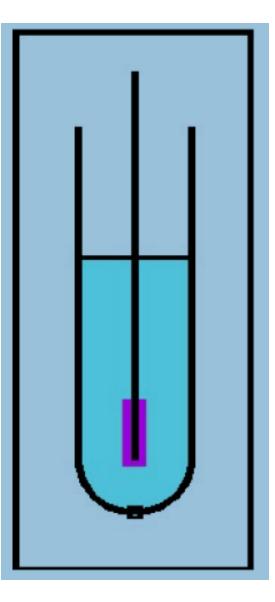
Probes – Reference vs Indicator

- Reference Electrode:
 - A reference electrode is an electrode which has a stable and well-known electrode potential.
 - Its high stability is achieved by employing the redox system, which must contain saturated concentrations in each of the
 participating solutions of the reaction.
 - For the measured potential to have meaning in this context, the reference electrode must be constructed so that its composition is fixed, and its response is stable over time, with observed changes in measured potential due solely to changes in analyte concentration.
 - Three types of reference electrodes:
 - Calomel: (Mercury/ Mercuric Chloride)
 - Silver/Silver Chloride
 - Safe and good temperature tolerance
 - Reversible reaction permits electro flow via ion carriers
 - Redox-Coupled Half Cell: (Platinum/Iodide Electrolyte Coupling)

$Ag^{0} \leftrightarrow Ag^{+} + e^{-}$

Probes – Reference vs Indicator

- Reference Electrode:
 - The silver reference wire MUST be coated with silver chloride.
 - Potassium chloride will dissolve the AgCl, thus it must be saturated in AgCl to prevent this.
 - lonic travel is via a ceramic frit or "junction" and allow flow
 - Some have ceramic plug frit, annular porous ring or ground glass joint.



Probes – Reference vs Indicator

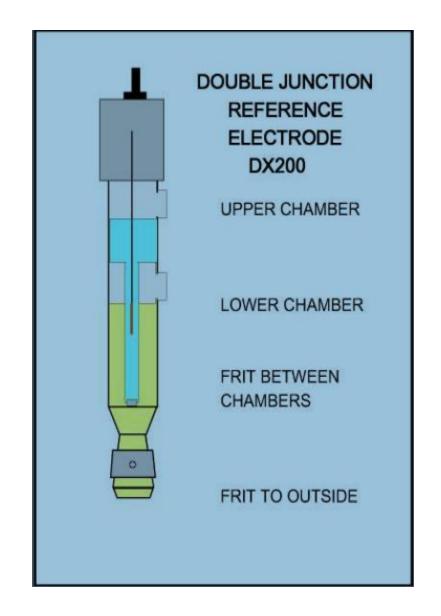
• Reference Electrode: Double

Junction

Double Junction mean the electrolyte in the lower chamber may be difference from that in the upper chamber and perhaps less reactive to sample constituents.

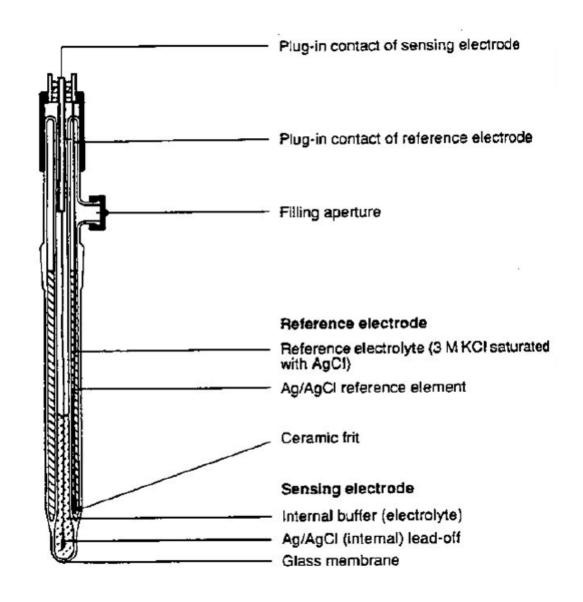
Typical alternate electrolytes might be the following.

3 M KCl 1 M NaCl KNO_3 LiCl in EtOH LiCl in glacial acetic acid $KClO_4$ in glacial acetic acid

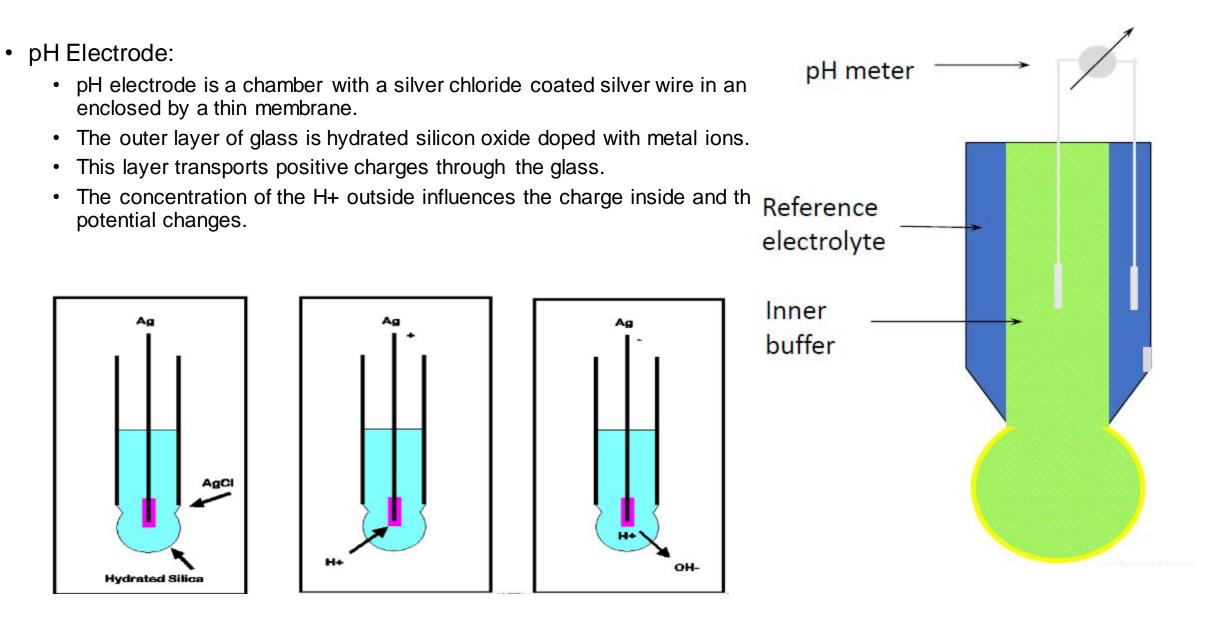


Probes

 Typical Combination pH Electrode



Probes



OTHER ANALYSES



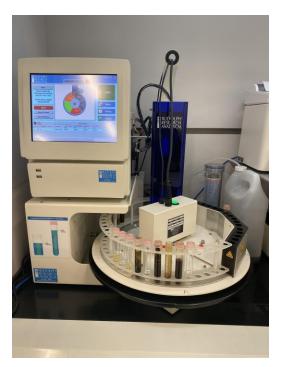
Characterization of all aspect of a sample

Total Sulfur/Nitrogen Analyzer Total Chloride Density Metal Content Viscosity



Applications cover all analyses

TNSA Densitometer ICP and ICP-MS Viscometer XRD XRF







Total Sulfur & Nitrogen Analyzers

$$SO_{2} + hv \rightarrow SO_{2}^{*}$$

$$SO_{2}^{*} \rightarrow SO_{2} + hv$$

$$R - S + O_{2} \xrightarrow{1050^{\circ}C} CO_{2} + SO_{2} + H_{2}O$$

$$R - N + O_{2} \rightarrow NO + CO_{2} + H_{2}O$$

$$NO + O_{3} \rightarrow NO_{2}^{*}$$

$$NO_{2}^{*} \rightarrow NO_{2} + h\gamma$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

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$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

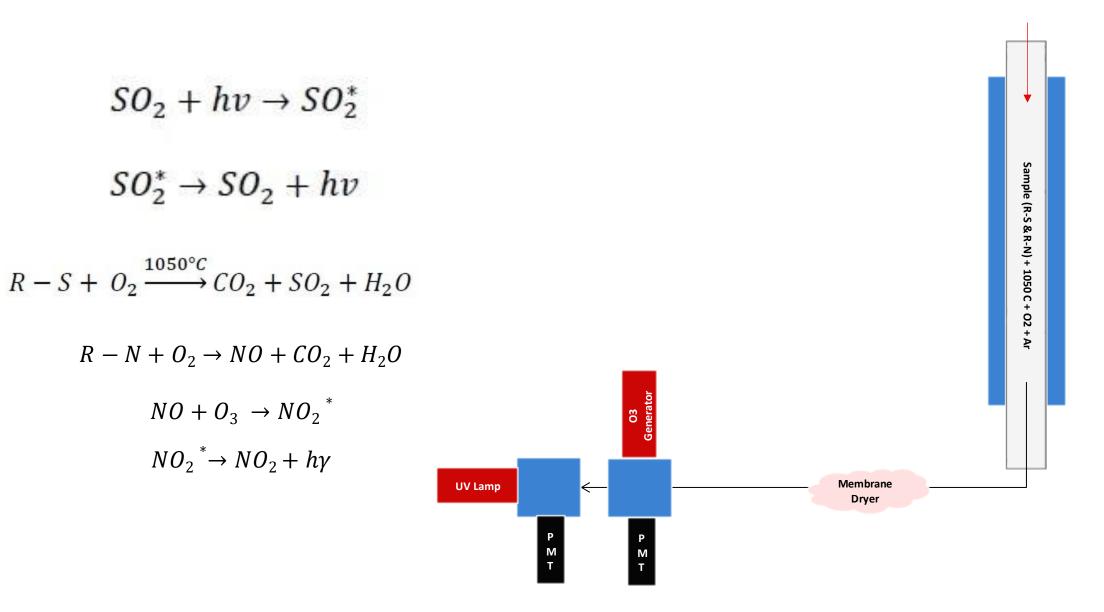
$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

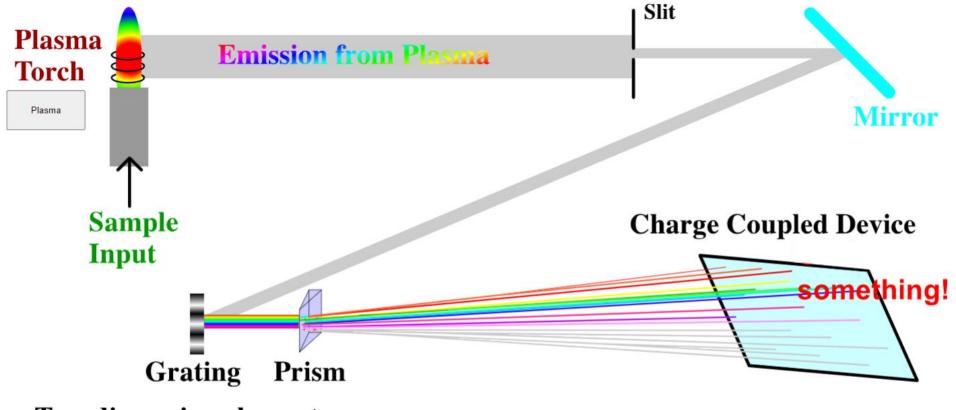
$$Sample (N-S) + 1050 C + O2 + Ar$$

$$Sample (N-S) + 1050 C + O2 + Ar$$

Total Sulfur & Nitrogen Analyzers



Inductively Coupled Plasma



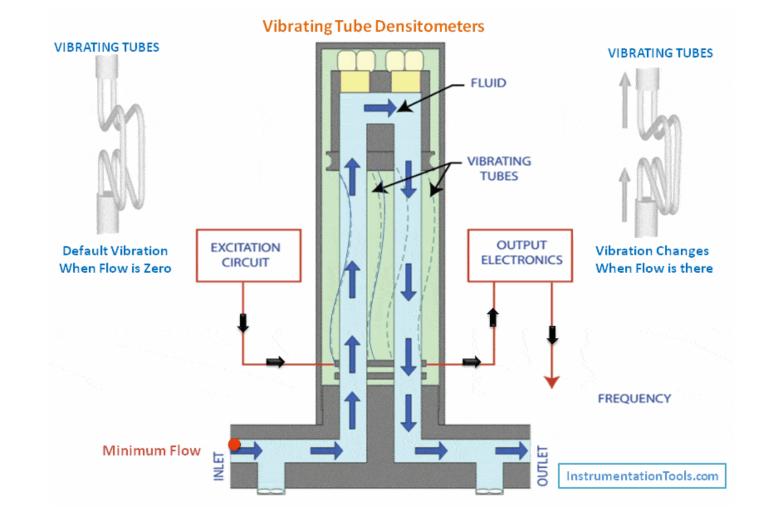
Two dispersion elements

						Fe
Primary λ , nm	327	294	291	285	267	238
Secondary λ	625	485	509	411	534	438
Tertiary λ	450	602	398	591	433	
Quaternary λ		403				

Many of the dispersed light rays shown here are in the ultraviolet and, therefore, actually invisible.

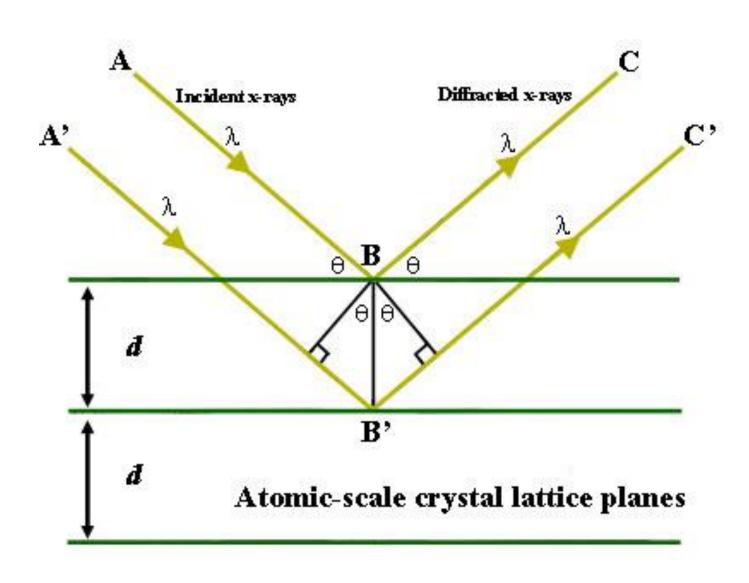
Densitometer Theory

 https://instrumentationtools.
 com/densitometers-workingprinciple/

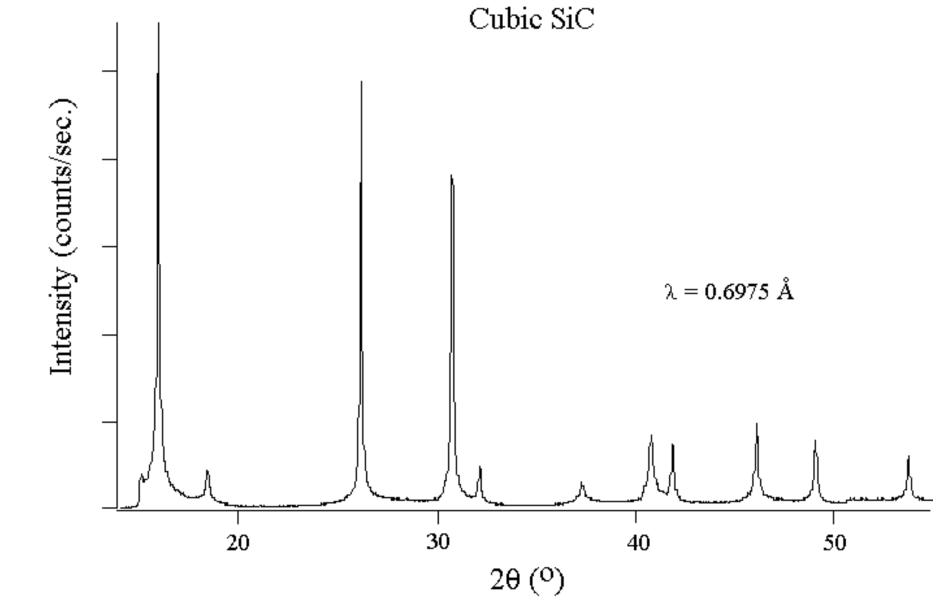


X-Ray Diffraction (XRD)
•Defined by Bragg's Law

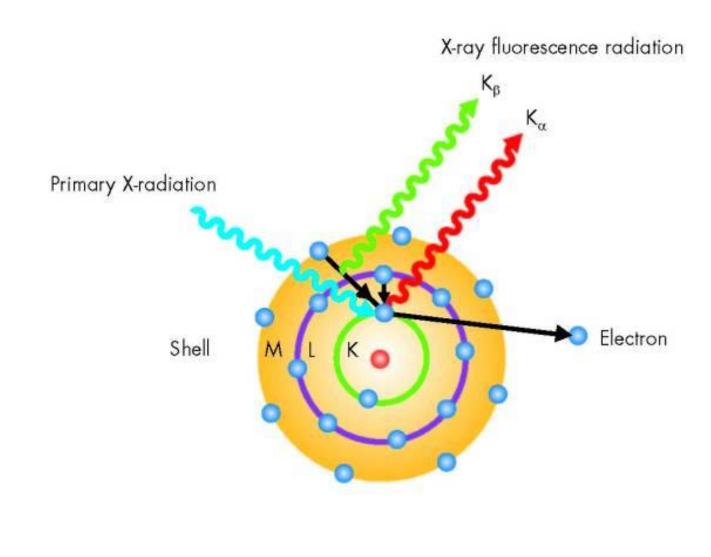
• $n\lambda = 2d \sin\Theta$



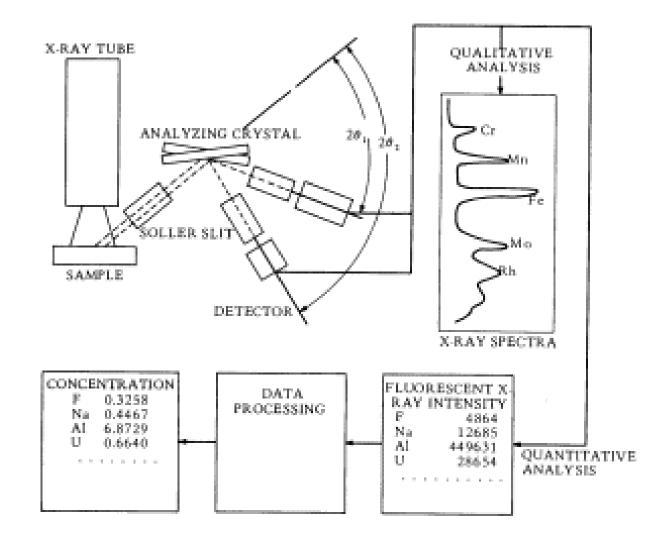
Example of XRD result



X-Ray Fluorescence (XRF)



X-Ray Fluorescence (XRF)



TYPICAL METHOD SOURCES

ASTM: American Society for Testing & Materials

IP: Institute of Petroleum, London

UOP: Universal Oil Products

Gas Processors Association

Open literature

Vendor technology

Documented Modifications

ASTM D2163-14 Standard Test Method for Determination of Hydrocarbons in Liquefied Petroleum (LP) Gases and Propane/Propene Mixtures by Gas Chromatography

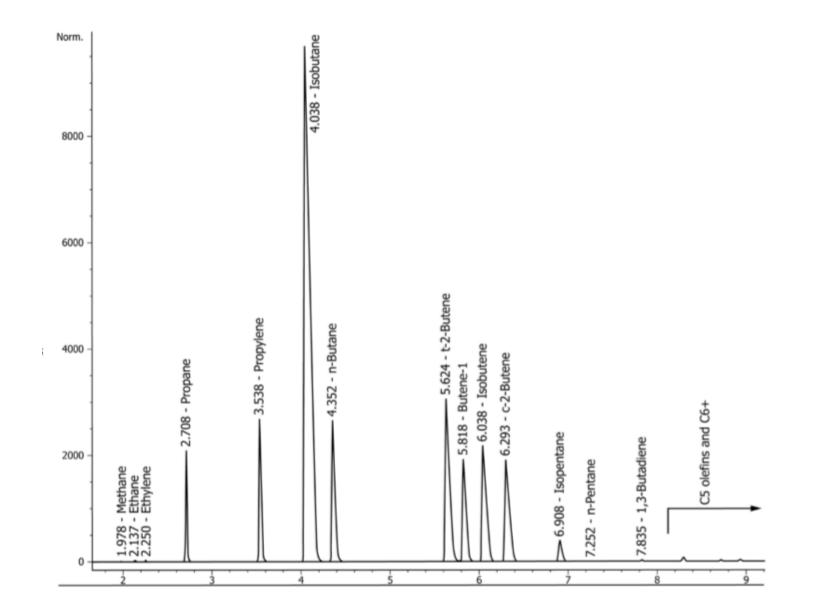
- 1. Scope
- 1.1 This test method covers the quantitative determination of individual hydrocarbons in liquefied petroleum (LP) gases and mixtures of propane and propene, excluding high-purity propene in the range of C1 to C5. Component concentrations are determined in the range of 0.01 to 100 volume percent.
- 1.2 This test method does not fully determine hydrocarbons heavier than C5 and nonhydrocarbon materials, and additional tests may be necessary to fully characterize an LPG sample.

ASTM D2163-14 Standard Test Method for Determination of Hydrocarbons in Liquefied Petroleum (LP) Gases and Propane/Propene Mixtures by Gas Chromatography

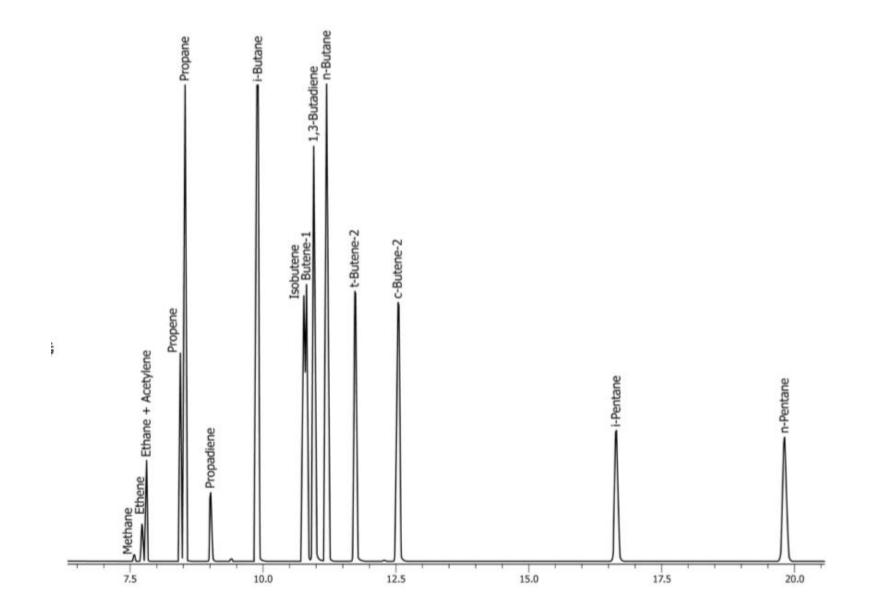
- Summary of Test Method
 - An LPG sample is analyzed via either liquid or gas sampling valves by gas chromatography and compared to corresponding components separated under identical operating conditions from a reference standard mixture of known composition or from use of pure hydrocarbons. The chromatogram of the sample is interpreted by comparing peak retention times and areas with those obtained for the reference standard mixture or pure hydrocarbons.
- Apparatus
 - Gas Chromatograph
 - Detector FID
 - Liquid Sampling Valve (recommended)
 - Gas Sampling Valve



Example Chromatogram Using the PLOT Column



Example Chromatogram Using the Dimethylpolysiloxane Column



Theoretical Mass Relative Response Factors

Component	RRFi	Component	RRF _i 0.874
Methane	1.0	1-butene	
Ethane	0.937	2-methylpropene (isobutene)	0.874
Ethene (Ethylene)	0.874	cis-2-butene	0.874
Propane	0.916	2,2-dimethyl propane (neopentane)	0.895
Propene (Propylene)	0.874	2-methyl butane (isopentane)	0.899
Cyclopropane	0.874	propyne (methyl acetylene)	0.834
2-Methylpropane (Isobutane)	0.906	cyclopentane	0.849
Ethyne (Acetylene)	0.813	n-pentane	0.899
Propadiene	0.834	1,3-butadiene	0.843
Butane	0.906	$C_5^{=}/C_6^{+}$ composite (backflush only)	0.885
Trans-2-Butene	0.874	$>nC_5$ ($C_5^{=}$ and heavier)	0.885

^ARF values obtained from Test Method D6729.

All response factors are relative to that of methane according to the following equation:

 $RRF_i = (MW_i/NC_i) \times (1/MW_{methane})$

where:

- RRF_i = relative response factor of each component with respect to methane,
- MW_i = the molecular weight of the component,
- NC_i = the number of carbon atoms in the component molecule, and
- $MW_{methane}$ = the molecular weight of methane.

Quality Control and Statistical Quality Assurance



What is SQC? Why do we do SQC?

•There were three degrees of untruth – a fib, a lie, and statistics

Charles Dilke



Measure each test, each method, each instrument, each analyst

լլլ

Measure overall lab performance

ישדי

How is an instrument behaving (Is it out of control?)



SQC can provide a deep insight into an issue

Yet, carelessness or duplicity can generate misleading results

What is SQC? Why do we do SQC?

SQC uses three scientific techniques

- sampling inspection
- analysis of the data
- control charting

Can be divided into following three broad categories

- descriptive statistics
- statistical process control
- acceptance sampling

QA/QC

Define Quality for Your Laboratory

- What does quality mean to your customers?
- Who are your customers?
- Precision of results

How good is good enough?

- Accuracy of 99%
 - 10 errors per 1,000 test
 - Typical lab analyze >200,000 test a year = 2,000 ERRORS!
- Accuracy of 99.99%
 - Typical lab analyze >200,000 test a year = **20 ERRORS**

Quality



• Our customer define what quality is

What is Quality?

- Has the notion of value, not just efficiency
- Difference between **efficiency** and **effectiveness** is the difference between **knowledge** and **wisdom**
- Directed toward what you want, not toward what you don't want

Peter Druker

- There is a difference between doing things right and doing the right things
 Doing things right Management
 Doing the right things Leadership
- Efficiency is doing things right, which is irrelevant if you are not doing the right things



QA/QC

Elements of Laboratory Quality	Test Quality
	Sample Quality
	Analyst Competence
	Customer Satisfaction
	Documentation
	Reputation
Path of	
Path of	Sampling
Path of	Sampling Laboratory Environment
Path of Workflow	
	Laboratory Environment
	Laboratory Environment Quality Control Procedures
	Laboratory Environment Quality Control Procedures Communications

QA/QC - Limits

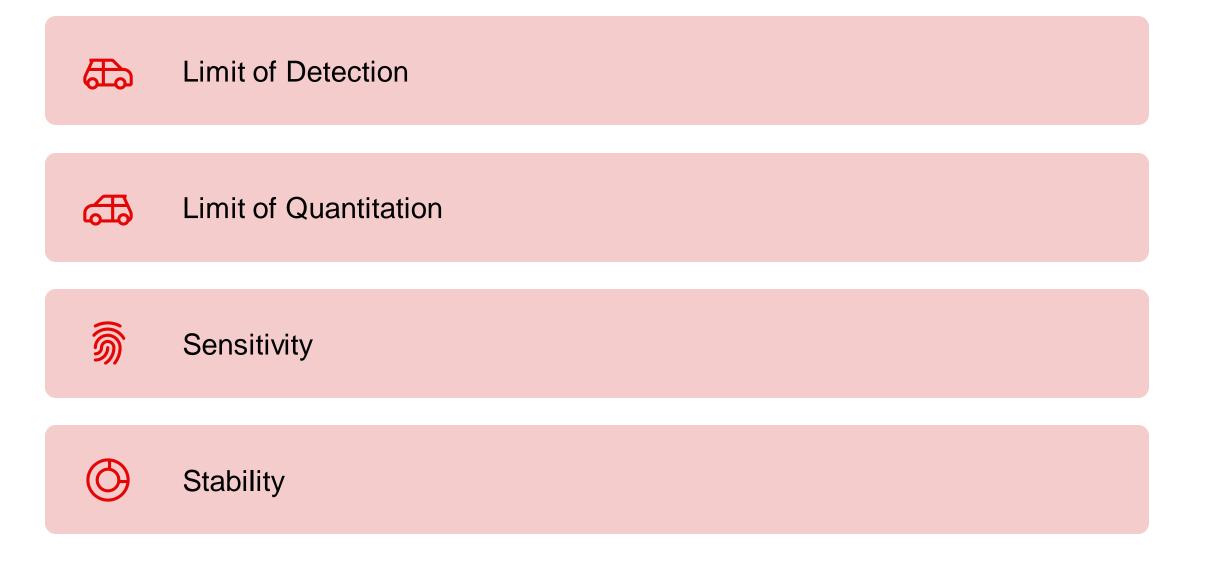
Limit of Detection

- The lowest amount of analyte in a sample which can be detected but not necessarily quantitated as an exact value
- 1:3 S/N ratio

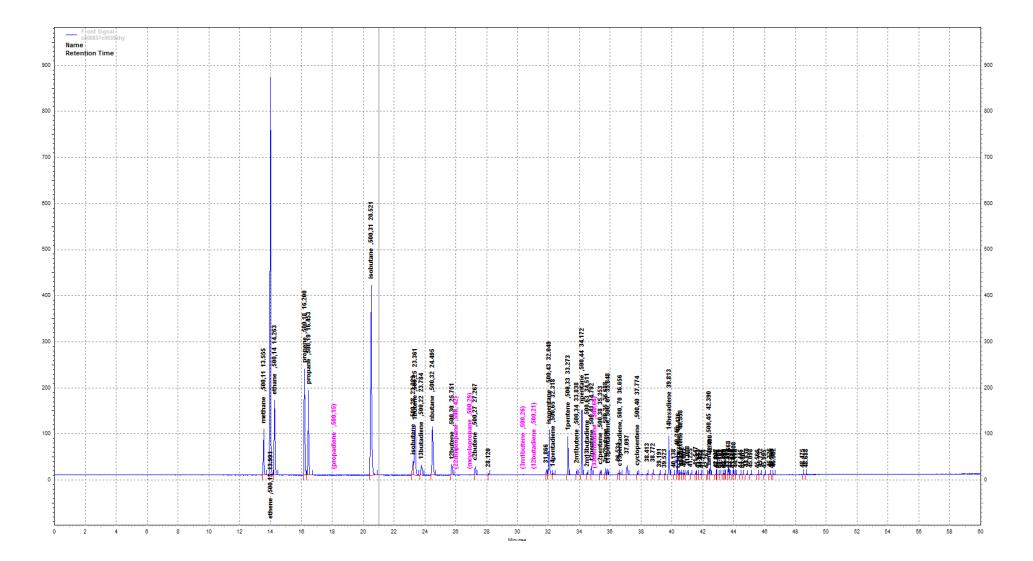
Limit of Quantitation

- The concentration at which quantitative results can be reported with a high degree of confidence
- 1:10 S/N ratio

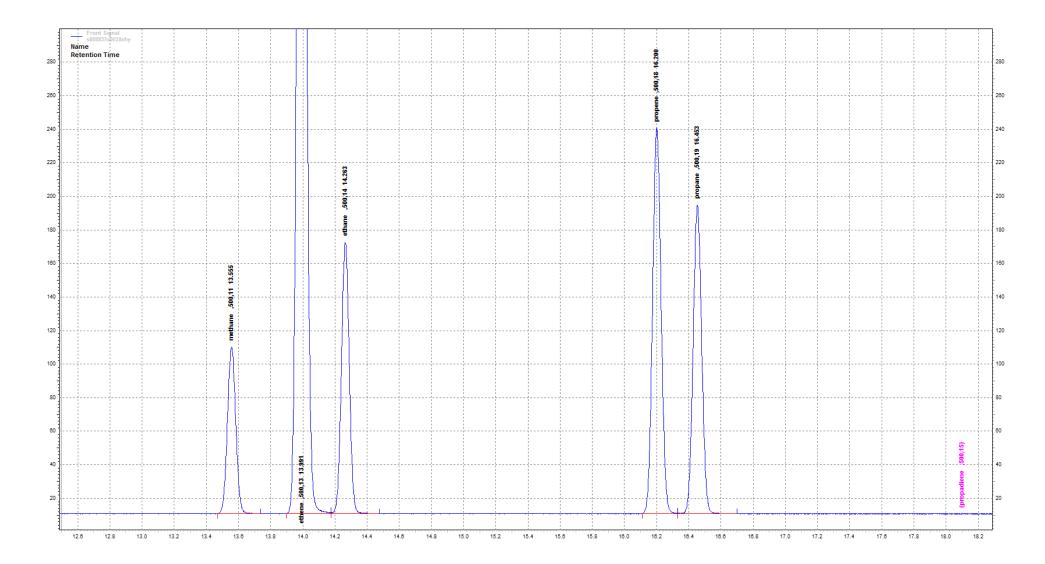
QA/QC - Instrument Performance Measures



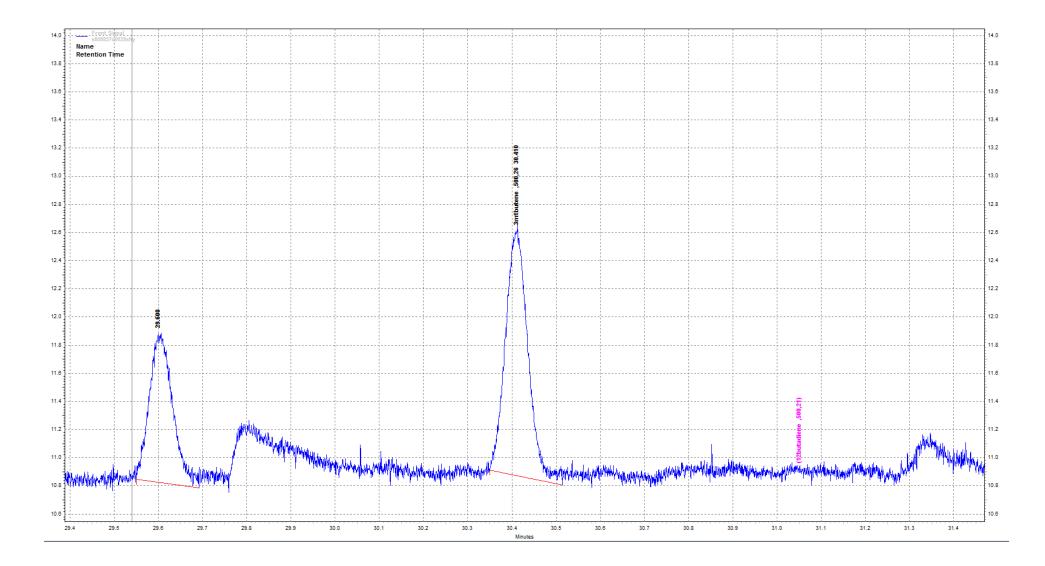
Full Chromatogram



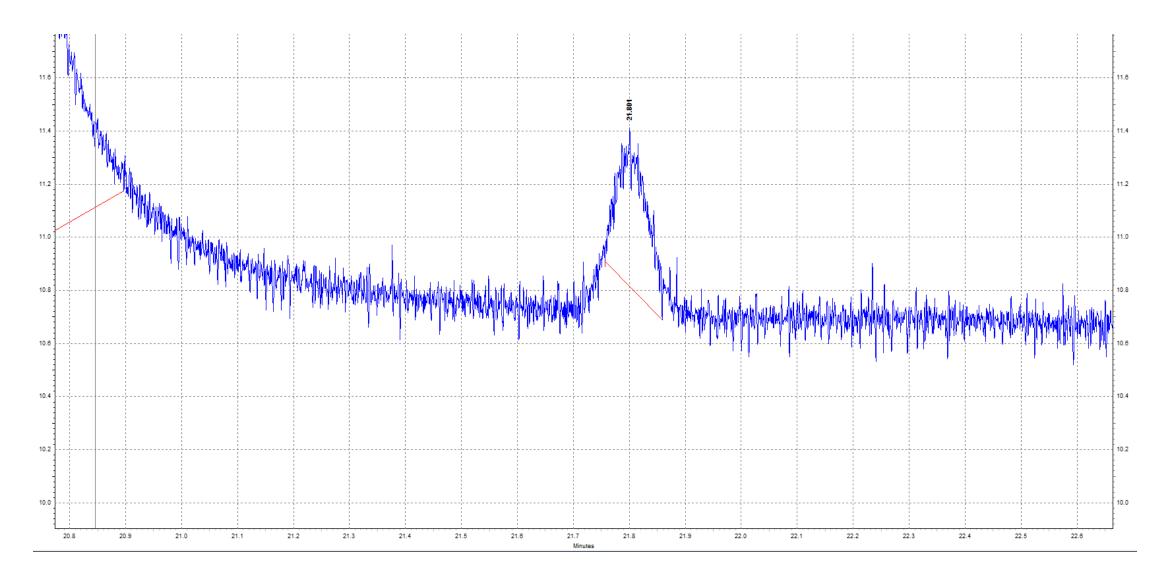
Expanded View



Quantitation Limit

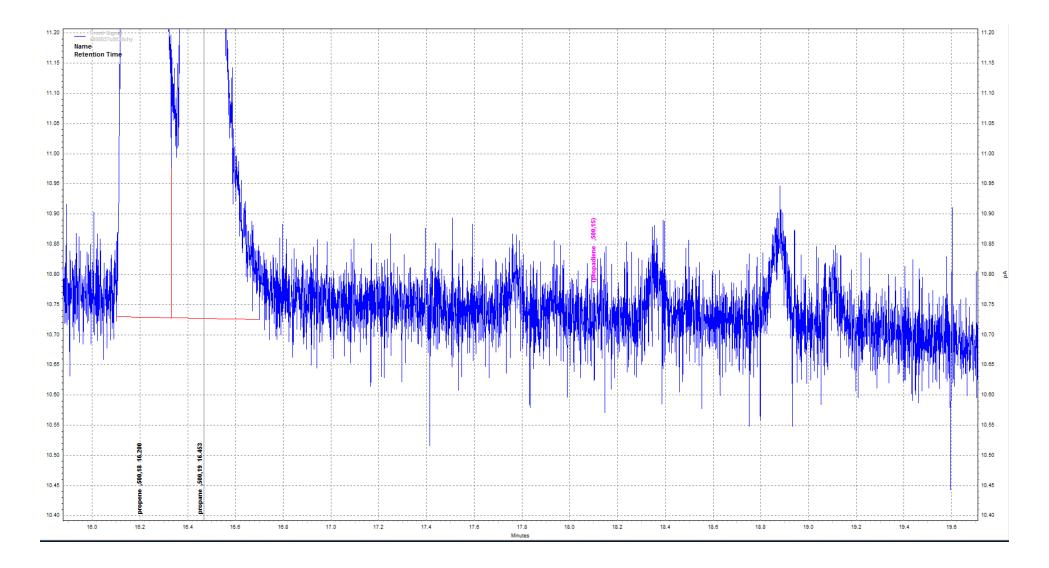


Detection Limit



154

Below Detection Limit



155

QA/QC - Standards

Commercial Suppliers

- Very expensive; slow delivery
- Quality variable

Home Made Blends

- Customize as needed
- Dilute/Modify commercial references
- Cheaper than commercial

Working Samples

- Process Origins; contains typical impurities
- Must be stable; unreactive
- Good for trending
- Cheap

QA/QC - Calibration

Establish a mathematical function which describes the dependency of the system's parameter on the measured value

Concentration

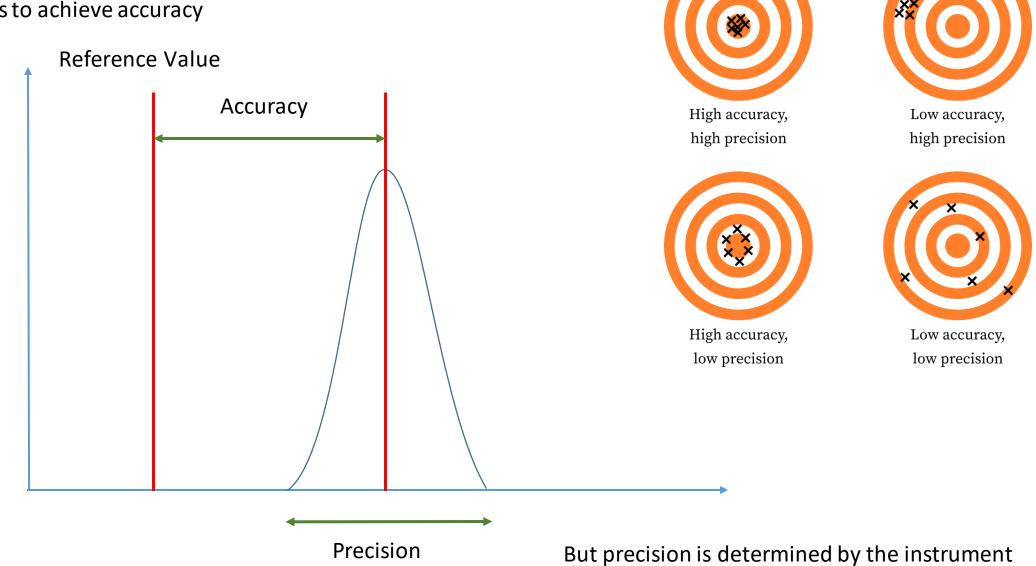
 Normalization vs External Standard

Gain statistical information of the analytical system

- Sensitivity
- Precision
- Accuracy

Accuracy vs Precision

Calibration adjusts to achieve accuracy



Repeatability and Reproducibility for Oxygenates (mg/kg)

NOTE 1—Where: X = the average individual oxygenate concentration of two results in mg/kg.

Analyte	Repeatability Limit (r)	Reproducibility Limit (R)
Acetone†	(0.1821)(X ^{0.5985})	(0.4424)(X ^{0.5985})
Acetyaldehyde	0.2595(X + 0.0001) ^{0.595}	1.0439(X + 0.0001) ^{0.595}
Diethyl Ether	0.1869(X + 0.0001) ^{0.5981}	0.5966(X + 0.0001) ^{0.5981}
Dimethyl Ether	0.05321(X + 0.0001) ^{0.9273}	0.2784(X + 0.0001) ^{0.9273}
DIPE	0.1188(X-0.6566) ^{0.5889}	0.5219(X-0.6566) ^{0.5889}
ETBE	(0.06778)(X ^{0.8512})	(0.3613)(X ^{0.8512})
Ethanol	$0.1626(X + 0.0001)^{0.7649}$	0.6808(X + 0.0001) ^{0.7649}
Iso-Propanol	(0.2458)(X ^{0.5108})	(1.1222)(X ^{0.5108})
MEK	(0.2009)(X ^{0.5094})	(0.7171)(X ^{0.5094})
Methanol	(0.2870)(X ^{0.4887})	(1.9695)(X ^{0.4887})
МТВЕ	(0.1261)(X ^{0.6368})	(0.2861)(X ^{0.9442})
n-Butanol	(0.1179)(X ^{0.9278})	(0.3890)(X ^{0.9278})
Sec-Butanol	(0.1063)(X ^{0.8057})	(0.5578)(X ^{0.8057})
TAME	0.2812(X + 0.0001) ^{0.4011}	0.9946(X + 0.0001) ^{0.4011}

†Editorially corrected.

Risk of being out of tolerance

Cost of calibration

Accuracy requirements

Experience with equipment and methods

Manufacturer recommendations

QA/QC - Common Sources of Analytical Error

Temperature and humidity variations
Weighing errors
Sampling errors
Instrument precision
Calibration standard error
Rounding error
Human error
Contamination
Reading error
Non-linearity
Flow rate variations

Flow rate variations

SOURCES OF ERRORS

Systematic Errors

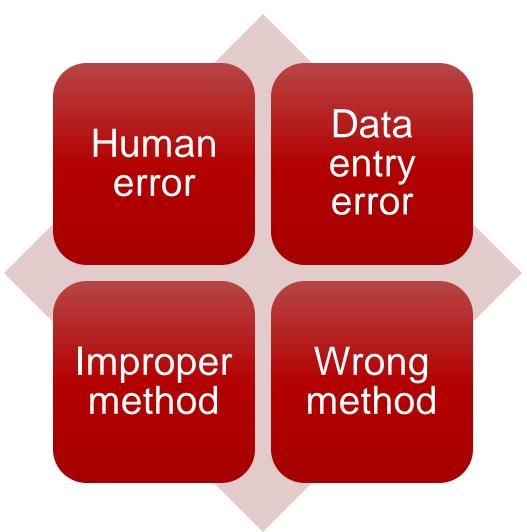
- Method limits
- Bias
- Mathematical correction

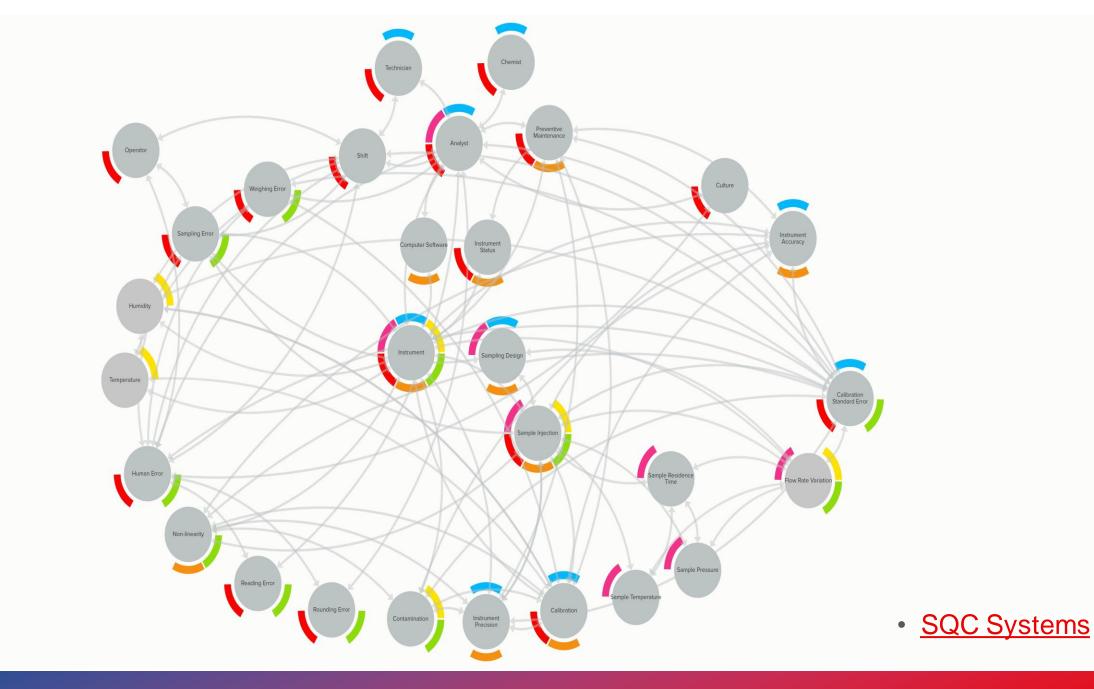
Random Errors

Normal variations

Statistical

SOURCES OF ERRORS - MISTAKES







---- Opposite

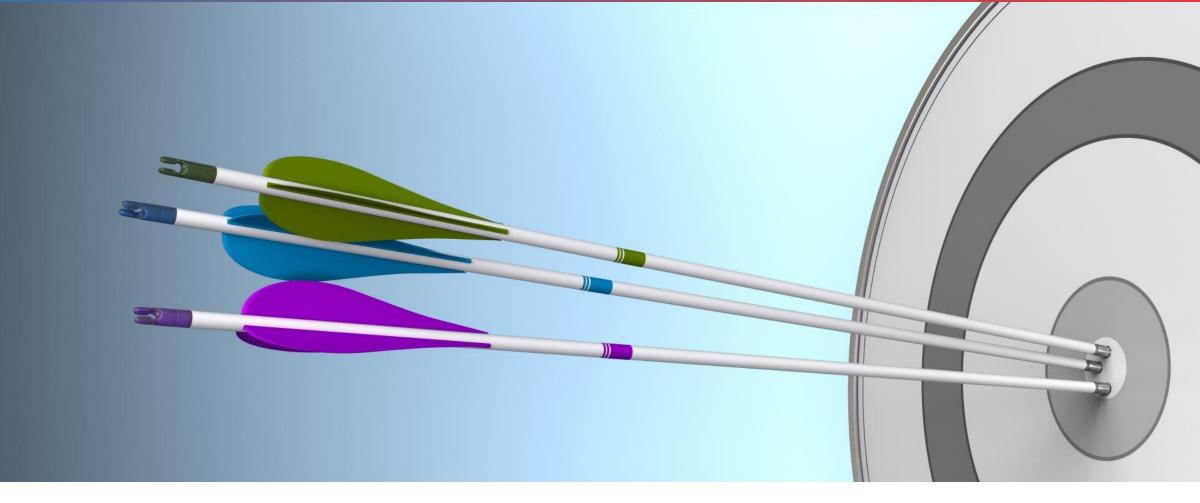
Calibration

EnvironmentError

Instrument

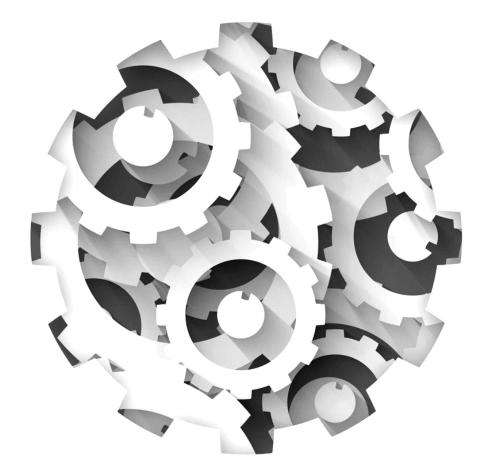
Personnel

Sample

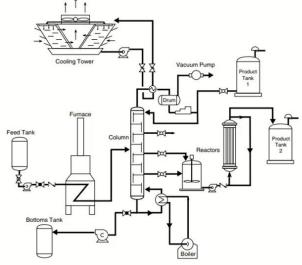


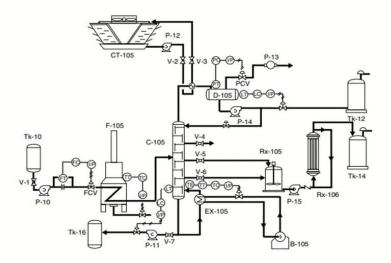
QA/QC – Customer Satisfaction

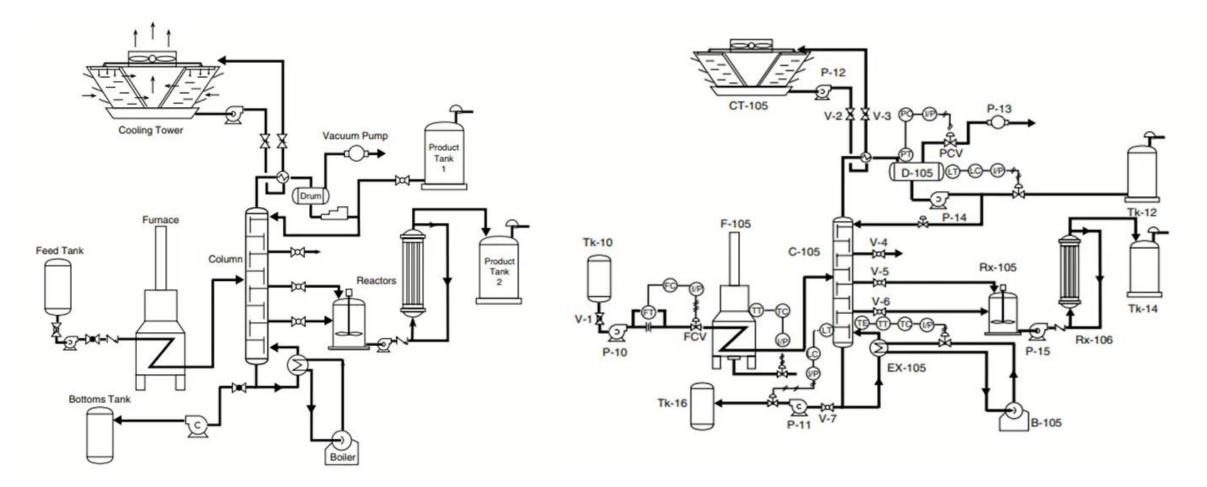
Meets or exceeds customer requirements

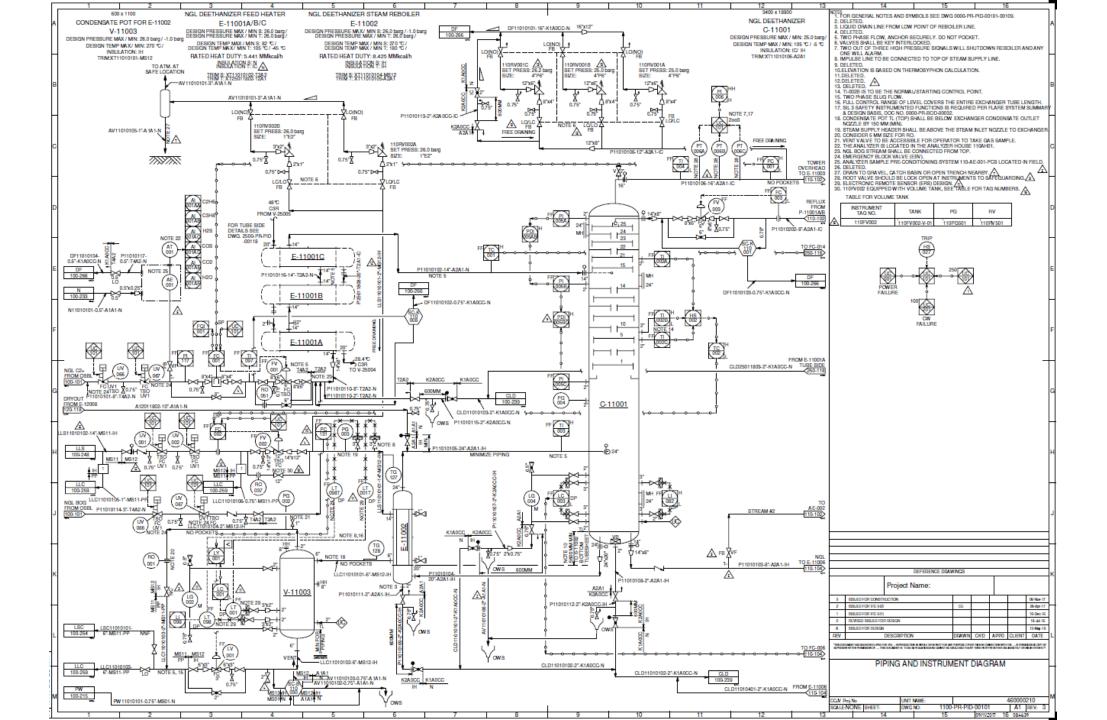


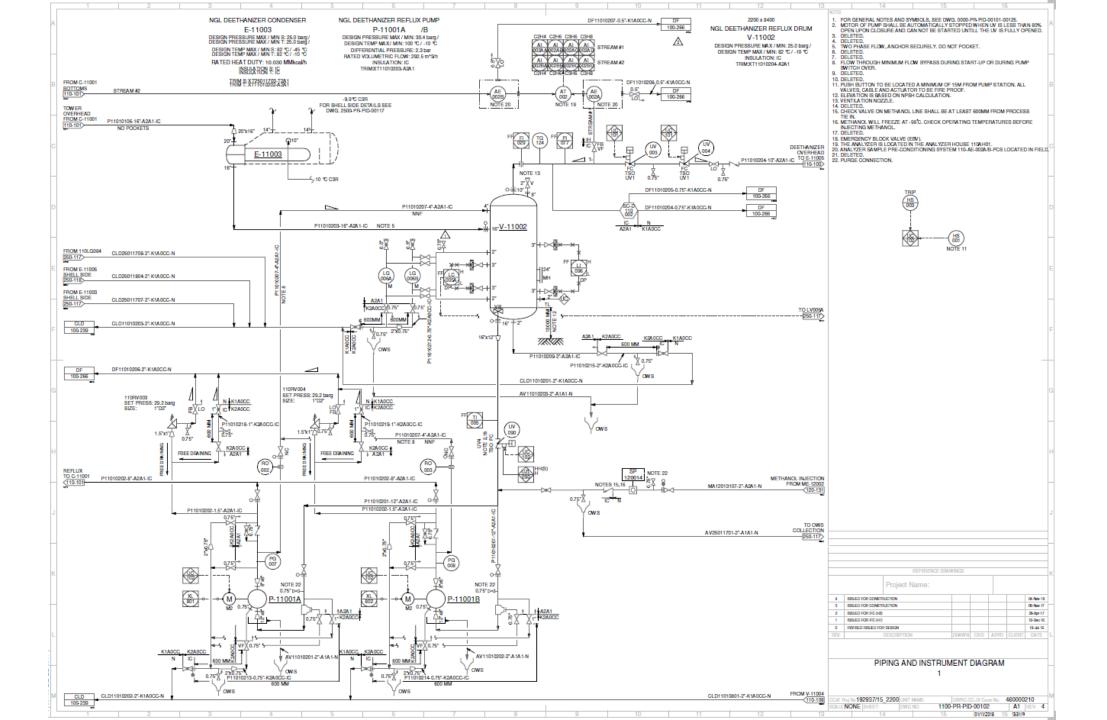
- PFD Process Flow Diagrams
 - The process flow diagram is also known as "flow sheet" is the first document that enables a clear understanding of the process to be controlled. It shows only the major equipment without details.
- P&ID Piping and Instrumentation Diagrams
 - A Piping and Instrument or instrumentation drawing comprises more details than a PFD. It is developed based on information from process flow diagrams, which is developed based on a Block flow Diagram.
 - A P&ID drawing includes both major and minor details of the process plant.

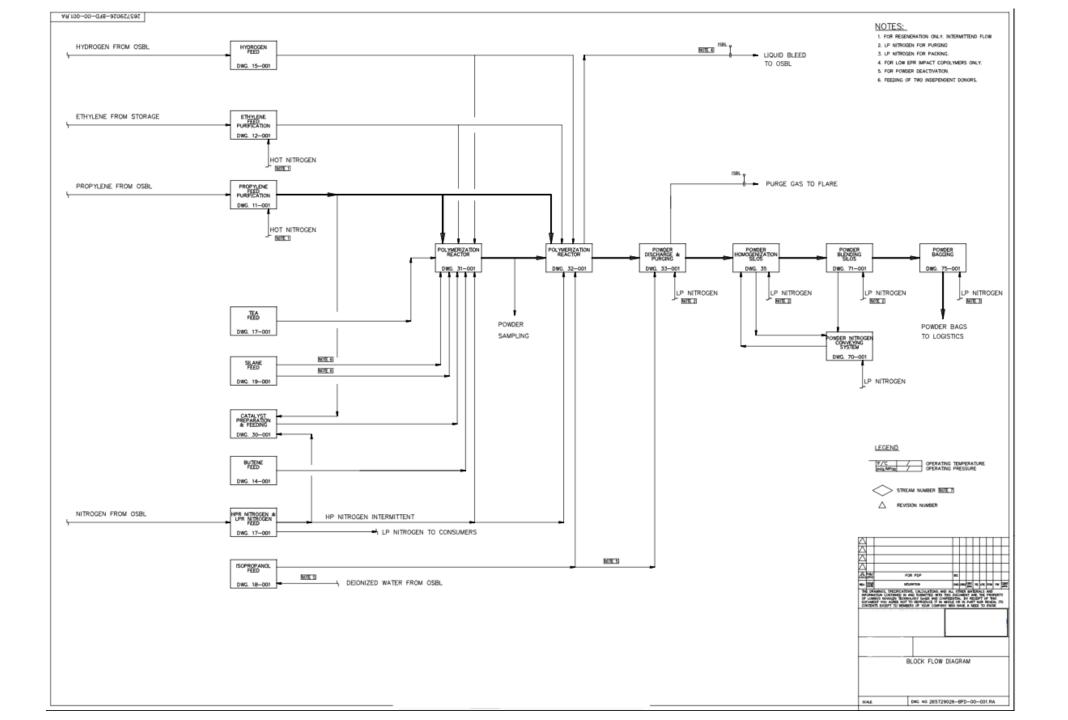




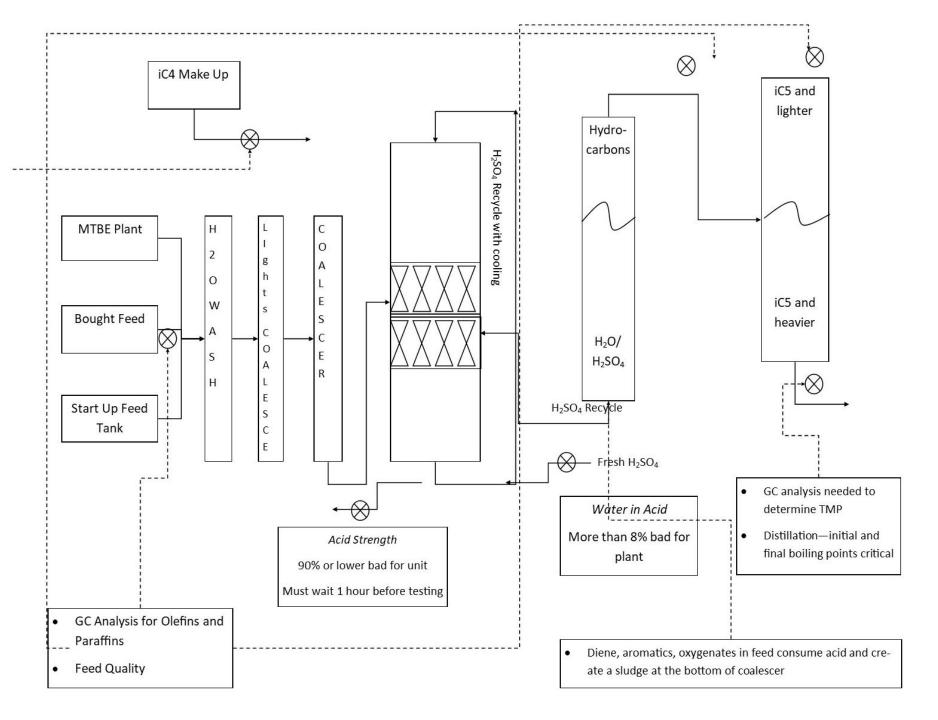




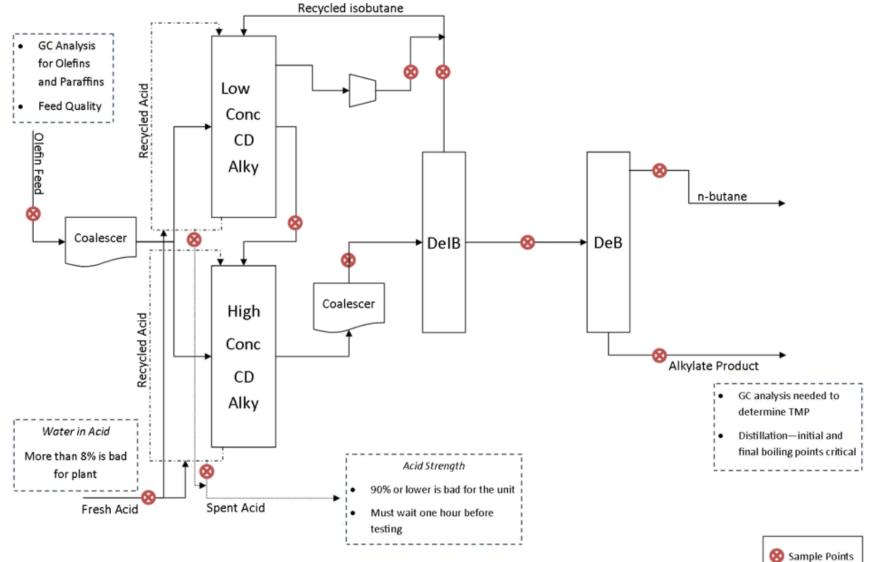




Flow Diagram



Simplified Flow Diagram

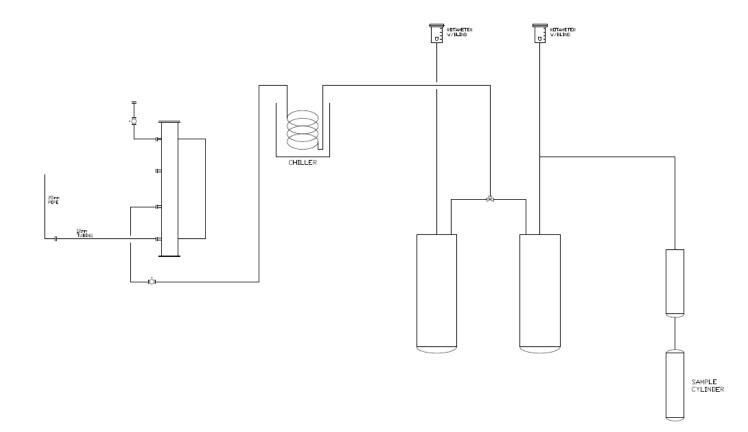


173



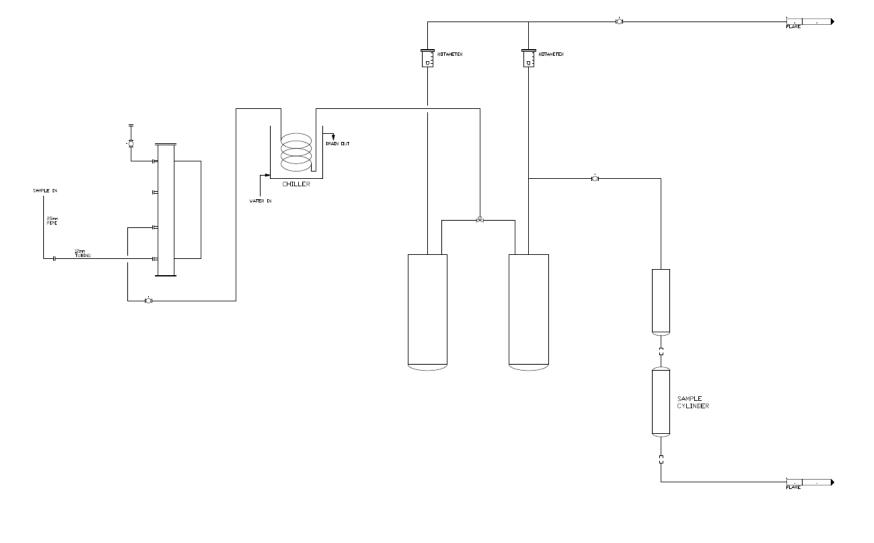


Sample Point Design

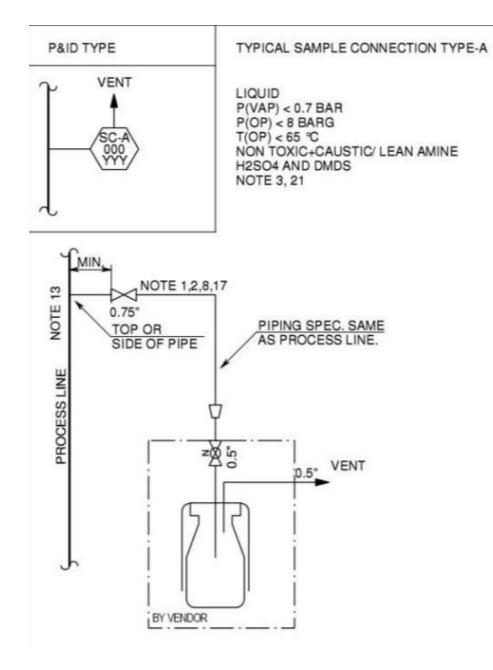


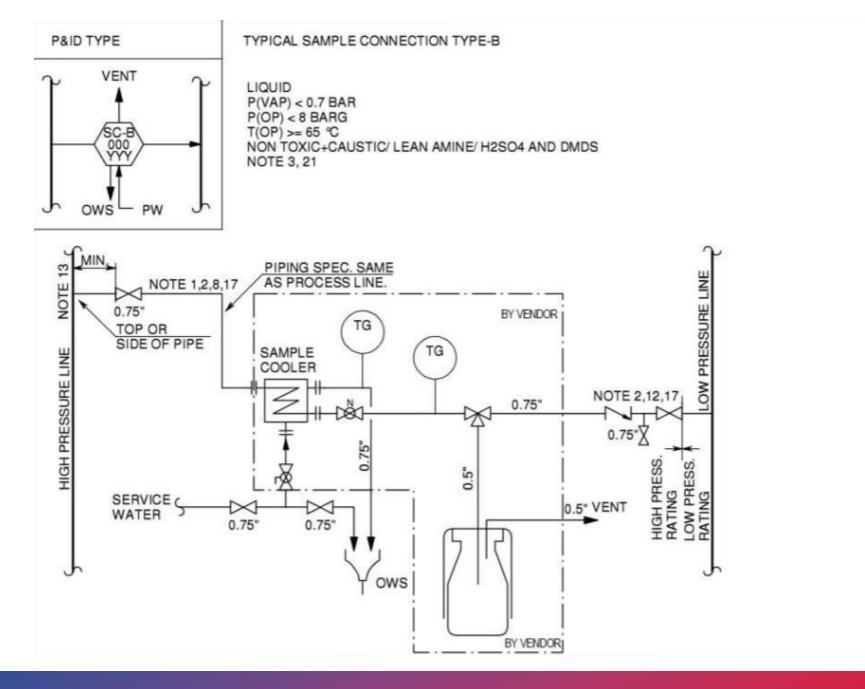


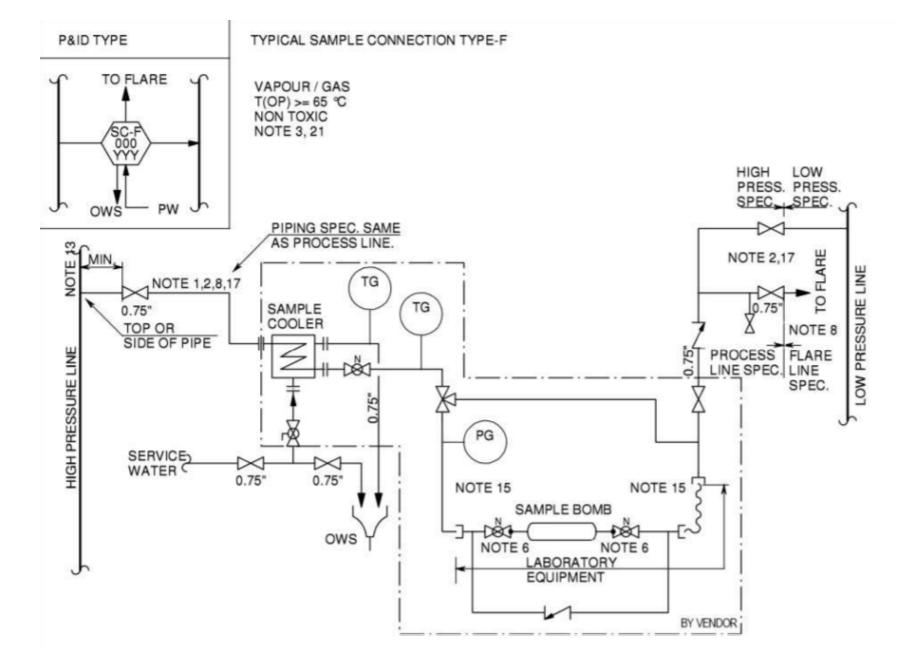
Sample Point Design



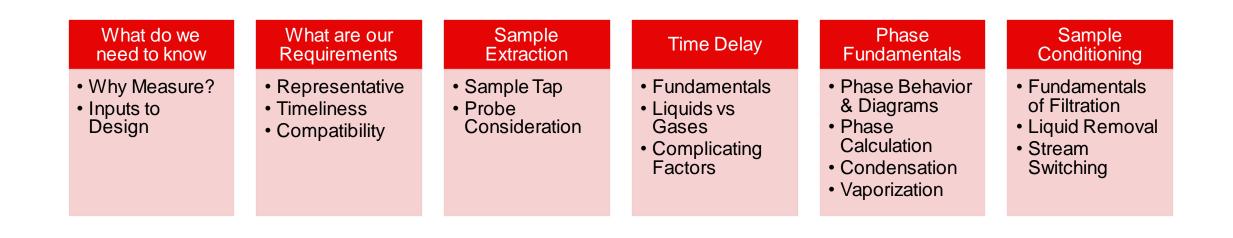
HEATER EFFLUENT SAMPLE POINT PROPOSED







Sampling System Fundamental



Why Measure?

Accounting / Custody Transfer

- Accuracy is critical
- Speed not important

Environmental Compliance

- Regulatory
- Accuracy and Validation is Critical

Direct Process Control

• Specific and Reliable

Process Monitoring

Reliable

Process Safety

- Speed and Accuracy
- Reliable
- Critical Alarms

Requirements

Representative

- Must be useful for its intended purpose
- Must provide the information needed to make decisions

Speed of Response

- Must respond quickly enough to provide the information needed to make a decision
- Must be able to achieve the goal of the analysis

Compatibility

- Material of construction can handle the process fluid
- Must be design to handle pressure, temperature, flow rates and other physical parameters

Summary in three points

Chemists

 Analyze the samples they receive to the best ability of the analytical equipment available to them

Engineers

• Utilize the analytical results provided to them to alter, improve, or change the process

Scientists

We are all scientists, but we focus of different aspect of science



Any final questions or comments?

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